



Effects of Malaria on Selected Liver Function Profiles of Children in Port-Harcourt, Rivers State, Nigeria

G. N. Wokem^{1*}, E. Nnadi², O. Azuonwu¹ and A. Okafor¹

¹*Department of Medical Laboratory Science, Rivers State University, Port Harcourt, Rivers State, Nigeria.*

²*Alvan Ikokwu Collage of Education, Owerri, Imo State, Nigeria.*

Authors' contributions

This work was carried out in collaboration between all authors. Author GNW designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors EN and OA managed the analyses of the study. Author AO managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJTDH/2018/42503

Editor(s):

(1) Francis Hombhanje, Centre for Health Research and Diagnostics, Divine Word University, Papua New Guinea.

Reviewers:

(1) Mahmud Ali Umar, Kano University of Science and Technology, Nigeria.

(2) Umar Adam Katsayal, Ahmadu Bello University, Nigeria.

(3) Kamgain Mawabo Lugarde, District Hospital of Deido, India.

Complete Peer review History: <http://www.sciencedomain.org/review-history/25641>

Original Research Article

Received 4th May 2018
Accepted 17th July 2018
Published 23rd July 2018

ABSTRACT

A cross sectional investigation of the effects of malaria infection on selected liver function profiles of children in Port Harcourt, Rivers State, Nigeria, was conducted. Exactly 1000 randomly selected children aged 1-10 years were involved after institutional ethical clearance and informed consent from their parents were obtained. Exactly 694 of the children, who were malaria positive represented the test group while 306 children apparently healthy, formed the control group. About 10 ml of blood was collected from each child through venopuncture with hypodermic syringe; 4 ml was dispensed into EDTA bottle for *Plasmodium* identification while 6ml each, was dispensed into lithium bottle for liver function tests. Thick and thin blood films were giemsa stained. Total bilirubin, conjugated bilirubin, unconjugated bilirubin, aspartate aminotransferase and albumin were assayed using standard biochemical techniques. Malaria prevalence was 69.4%; age group 1-5 years was

*Corresponding author: E-mail: g.ndrwokem@yahoo.com;

significantly ($P<0.05$) more infected (39.7%) than age group 6-10 years (29.7%). Total bilirubin, conjugated bilirubin, unconjugated bilirubin and aspartate aminotransferase were significantly elevated ($P<0.05$) in malaria subjects. There was a statistically significant ($P<0.5$) reduction of albumin levels in tests than in the controls. Among high malaria parasitaemic subjects, total bilirubin, conjugated bilirubin, unconjugated bilirubin, and aspartate aminotransferase were more elevated significantly ($P<0.05$) than those of low parasitaemic subjects. These significant changes in these parameters suggest that malaria parasitaemia has significant negative effects on the integrity and functions of the liver which may lead to mortality if ignored.

Keywords: Malaria; liver-function-profiles; effects; children; Port-Harcourt; Nigeria.

1. INTRODUCTION

Malaria is the most important parasitic infection in the world; it is very common in the tropical regions but can also occur rarely in temperate regions. Its transmission, morbidity and mortality are greatest in Africa where most deaths from malaria are among young children and pregnant women [1,2]. It accounts for 1 in 5 of all childhood death in Africa. Anaemia, low birth weight (LBW), abortions, stillbirths, growth/mental retardation and neurologic problems, are frequent complications of malaria and are known to have devastating consequences on the general population and these compromise the health and development of millions of children throughout the tropics [3,4,5].

The *Plasmodium* species commonly involved in human malaria are *Plasmodium falciparum*, *P. vivax*, *P. ovale*, and *P. malariae*. Some human cases of malaria have also been reported with *P. knowlesi*, monkey - malaria that occurs in certain forested areas of South East Asia [6,7]. Many species of female *Anopheles* mosquitoes transmit malaria parasite in different parts of the world and different species have varied vectorial potentials. Among the most competent malaria vectors are female *Anopheles gambiae* and *Anopheles funestus* which are dominant in Africa while female *Anopheles darlingi* is the major vector in the Amazon basin [8,9]. Children in endemic areas experience frequent episodes of malaria. Initial episodes are commonly severe and the majority of deaths from malaria occur in young children from 6 months of age in endemic areas. The prevalence of malaria among children has been reported in many parts of Nigeria and this is of great importance. Some others, [10,11], reported prevalence of 67.2% and 65.5% among nursery and primary school pupils in Ataba and Port Harcourt, Rivers State, Nigeria respectively while [12] recorded a prevalence rate of 80.5% in Ota, Ogun State. [13] also reported a prevalence rate of 80.4% in Aba, Abia State with Enugu State having the least malaria infection rate of

35.8%. However, Asia, Latin America and to a lesser extent Middle East and parts of Europe are also affected; about 91 countries of the world with 216 million cases are infected by this debilitating protozoan infection [14,15].

Plasmodium parasite interferes with the organs of the body namely brain, kidney, liver, lungs, central nervous system and spleen etcetera. Hepatic dysfunction was reported to be 7.15% by [16] at Minna, Niger State, Nigeria. This gives much concern about the need for timely diagnosis, early treatment and the application of effective preventive measures to avert the severe consequences, hence the need for a deeper insight into the problem [17]. According to [18], liver involvement in malaria seemed common in patients with severe malaria and may manifest as jaundice, that is raised serum bilirubin, hepatomegaly, elevated liver enzymes like aspartate aminotransaminase/ alanine transaminase; there may be a reduced level of serum albumin and prothrombin time may also be prolonged. Specific histopathological changes and cases with altered liver function tests, (fulminant hepatic failure and hepatic encephalopathy) have been reported [18]. Some researchers [19] also reported that malaria hepatitis is characterized by increased levels of transaminase levels to more than three times the upper limits of normal and alkaline phosphate. When bilirubin is then excreted from the body through the faeces and the elevated levels may indicate certain diseases. Bilirubin circulates in the blood stream in two forms; unconjugated (indirect) bilirubin which does not dissolve in water due to intra molecular hydrogen bonding. It is then bound to albumin and sent through the blood stream to the liver where it is changed into a soluble form (direct or conjugated). Conjugated (direct) dissolves in water because bilirubin is conjugated with glucuronic acid and is then made by the liver from indirect bilirubin much of which goes into the bile and out of the small intestine [20].

Studies on the interactions between malaria and liver enzymes have been carried out in other places like in Minna, Niger State and Edo State [16,21] but much work has not been done among children in study area. Also, in adults there could be many factors that can cause liver disease such as alcoholism [22,23] but children in the study area are not implicated to such; therefore, any derangement in the liver function tests may be attributed to malaria. This work is aimed at investigating the effect of malaria infection on liver enzyme function profiles in children.

2. MATERIALS AND METHODS

The study was carried out among children attending Palmars, Omega Children Hospital, Braithewait Memorial Specialist Hospital (BMSH) and Schools (Early Breed Group of Schools, St Francis Nursery and Primary School and Staff Nursery and Primary School) in Port Harcourt, Rivers State. Port Harcourt is situated at latitude 4°47'21''N and longitude 6°59'54''. One thousand (1000) children were included in this study. Six hundred and ninety four (694) children had malaria and were regarded as test group while 306 children who had no malaria were regarded as the control group.

2.1 Experimental Design

This is a cross sectional study where the subjects were randomly selected. The sample size was derived using the following formula [24]:

$$N = \frac{Z^2 (PQ)}{D^2}$$

Minimum sample size required for this study was calculated using the a reported prevalence of 65.6% from the same town [11]. The test population was 694 with 306 control, making up the number to 1000 subjects. The research work was started in April, 2011 and finished May, 2015; this is just a section of the work.

2.2 Blood Collection and Laboratory Assays

About 10 ml of blood samples were collected through the vein with disposable hypodermic syringe. About 4 ml of which was dispensed into ethylene diethyl tetracetic acid (EDTA) bottle for malaria parasite test while 6ml was used for Liver function tests using total bilirubin, conjugated bilirubin, unconjugated bilirubin, albumin and aspartate aminotransferase, as parameters.

The parameters assayed were total bilirubin using Jendrassik Groff by Mally, et al., [25]. It was used for the quantitative determination of total bilirubin in serum or plasma. Conjugated bilirubin was done using Groff technique by Martinek [26]; unconjugated bilirubin was determined by subtracting the direct bilirubin from the total bilirubin result. Aspartate aminotransferase estimation was done using enzymatic method by Reitman and Frankel [27] for its quantitative determination in serum while Bromocresol Green Method by Grant and Kackmser [28] technique was used for albumin qualitative estimation. Blood films preparation and *Plasmodium* identification were made according to the researcher [29]. Estimation of parasite density was done using quantitative method as described by Cheesbrough [29] where 1+ denotes low parasite density (1-10 parasites per 100 thick film fields), 2+ denotes moderate parasite density (11-100 parasites per 100 thick film fields) and 3+ denotes high parasite density (1-10 parasites per thick film fields).

2.3 Statistical Analysis

The data generated were statistically analyzed using statistical package for Social Science (SSS) version 21 and Mega Stat, one factor ANOVA. The results were expressed as mean, standard deviation, per cent, variance and P-value used equals to or less 0.05 as statistically significant. Independent Chi-square test for comparison of proportions was also used.

3. RESULTS

Table 1 shows that overall malaria prevalence was 69.4% (test) while age group 1 – 5years had 39.7%; age group 6- 10 years was 29.7%. There were statistically higher significant differences (P ≤ 0.05).

Table 2 shows the Mean Parasite Density per microlitre of blood and the corresponding liver function parameters. It was observed that parasite count of <1000 per microlitre of blood had lower values of total bilirubin (TB), conjugated bilirubin (CB), unconjugated bilirubin (UNB) and aspartate aminotransferase (AST), when compared with the parasite count of >1000 ≤ 9999 per microlitre of blood and >10000 per microlitre of blood. Parasite count of >1000 ≤ 9999 per microlitre of blood had lower values of total bilirubin (TB), conjugated bilirubin (CB), unconjugated bilirubin (UNB) and aspartate aminotransferase (AST) when compared with the parasite count of >10000 per microlitre of blood.

Table 1. Age related prevalence of malaria among the studied population

Age group (%) (Years)	Number examined	Number infected (Test group)	Uninfected (%) (Control)	X2(df)	P-value
1-5	584	397 (39.7%)	187 (18.7%)		< 0.05
6-10	416	297 (29.7%)	119 (11.9%)	964.311(1)	
Total	1000	694 (69.4%)	306 (30.6%)		

Legend: Mean = Control (30.6%) = % uninfected of 1000 subjects examined

Protein and albumin levels of parasite count of >10000 per microlitre of blood was lower than the levels in both <1000 and >1000≤9999 parasite count of per microlitre of blood.

Table 3 shows the Comparative Means (±SEM) of Liver Function Test Parameters of the Test and Control. It was observed that malaria infected subjects had significantly higher levels (P ≤ 0.05) of total bilirubin (TB), conjugated bilirubin (CB) and unconjugated bilirubin (UNB) and aspartate aminotransferase (AST) than the control group. Also there were significant lower levels (P ≤ 0.05) of albumin in malaria infected subjects than the control group.

Table 4 shows that there was a significant higher levels (P ≤ 0.05) of total bilirubin (TB), conjugated bilirubin (CB) and unconjugated bilirubin (UNB) and aspartate aminotransferase (AST), in malaria infected group than the control but there was no significant difference in levels (P ≥ 0.05) of albumin between the two groups. The Comparison between the Parameters in Low and High Densities showed a significant higher level (P ≤ 0.05) of total bilirubin (TB), conjugated bilirubin (CB) and unconjugated bilirubin (UNB) and aspartate aminotransferase (AST). There was no significant difference in level (P > 0.05) of albumin between the two groups.

4. DISCUSSION

The prevalence of malaria infection in the study area was 69.4%. The value is slightly higher 65.5% than [11] in Port Harcourt, Rivers State but lower when compared with 80.5% prevalence rate obtained by [12] at Ota, Ogun State, and 80.4% obtained by [13] at Aba, Abia State all in Nigeria. The result confirmed that children under the age of 5 years are mostly at risk which agrees with the results obtained by earlier researchers [11]. This may be probably due to immature immune system, low level of protection against mosquito bites, high exposure rates as well as nutritional factors [13].

In this study, there was an increase in bilirubin level in malaria infected subjects when compared with the control with a statistically significant difference (P ≤ 0.05). This agrees with the findings of [30,31]. Who suggested that hyperbilirubinaemia in malaria could be due to a number of causes like intravascular haemolysis of parasitized red blood cells and micro angiopathic haemolysis associated with disseminated intravascular coagulation. Unconjugated hyperbilirubinaemia is due to massive intravascular haemolysis whereas conjugated hyperbilirubinaemia is due to hepatocyte dysfunction and this is associated with raised transaminases.

Table 2. Mean parasite density/µl blood and the corresponding liver function parameters

Parameters	Parasite density <1000/µl (873.8 ± 30.44)	Parasite density >1000 ≤ 9999/µl (3248 ± 109.31)	Parasite density >10000/µl (24813.8 ± 877.22)
TB (µmol/l)	12.43 ± 0.16	15.40 ± 0.25	20.63 ± 0.63
CB (µmol/l)	4.54 ± 0.11	5.52 ± 0.15	7.03 ± 0.21
UNB (µmol/l)	8.06 ± 0.14	9.85 ± 0.22	13.06 ± 0.48
AST (iu/l)	14.73 ± 0.32	17.11 ± 0.11	24.03 ± 0.63
Albumin (g/l)	31.72 ± 0.41	30.61 ± 0.47	30.46 ± 0.46

Statistical significance: P < 0.05.

Legend: Low parasitaemia denotes parasite density of <1000/µl of blood

Moderate parasitaemia denotes parasite density of >1000 ≤ 9999/µl of blood

High parasitaemia denotes parasite density of >10000/µl

TB = Total bilirubin, CB = Conjugated bilirubin, UNB = Unconjugated bilirubin, AST = Aspartate aminotransferase

Table 3. Comparative means (\pm SEM) of liver function test parameters of the test and control groups from age 1 to 10 years

Parameters	Test	Control	P-value
	n=694	n=306	
TB (μ mol/l)	15.61 \pm 0.24	4.84 \pm 0.11	P < 0.05 = 5.48E- 137
CB (μ mol/l)	5.51 \pm 0.09	1.84 \pm 0.05	P < 0.05 = 3.57E- 109
UNB (μ mol/l)	10.16 \pm 0.18	2.99 \pm 0.83	P < 0.05 = 1.09E- 113
AST (iu/l)	18.20 \pm 0.32	5.64 \pm 0.11	P < 0.05 = 3.25E - 112
Albumin (g/l)	31.03 \pm 0.26	54.03 \pm 0.60	P < 0.05 = 2.41E - 218

Age Range 1-10 years

Legend: TB =Total bilirubin, CB = Conjugated bilirubin, UNB = Unconjugated bilirubin, AST = Aspartate aminotransferase, SEM = Standard error of the mean

Table 4. Comparison between the parameters in low, moderate and high densities

Parameters	1+	2+	3+	P-value
	n=298	n=199	n=197	
TB (μ mol/l)	12.43 \pm 0.16	15.40 \pm 0.25	20.63 \pm 0.63	P < 0.05 = 2.32E-42
CB (μ mol/l)	4.54 \pm 0.11	5.52 \pm 0.15	7.03 \pm 0.21	P < 0.05 = 1.19E-27
UNB (μ mol/l)	8.00 \pm 0.14	9.88 \pm 0.22	13.60 \pm 0.48	P < 0.05 = 1.60E-34
AST (iu/l)	14.73 \pm 0.32	17.11 \pm 0.49	24.03 \pm 0.63	P < 0.05 = 0.266
Albumin (g/l)	31.72 \pm 0.41	30.65 \pm 0.47	30.46 \pm 0.46	P > 0.05 = 0.0915

Legend: 1+ denotes low parasite density (1-10 parasites/100 thick film fields)

2+ denotes moderate parasite density (11-100 parasites/100 thick film field)

3+ denotes high parasite density (1-10 parasites/thick film field)

TB =Total bilirubin, CB = Conjugated bilirubin, UNB = Unconjugated bilirubin, AST = Aspartate aminotransferase.

In this study, most infected subjects showed unconjugated hyperbilirubinaemia which seemed to be due to haemolysis of peripheral parasitized red cells and impairment in bilirubin transport because of reticuloendothelial blockade and disturbance of hepatocyte microvilli which is a feature of falciparum malaria in agreement with [32]. There were also statistically significant ($P \leq 0.05$) higher values of total bilirubin and unconjugated bilirubin in moderate parasitaemia compared with low parasitaemia. Higher values of total bilirubin and unconjugated bilirubin were noticed in high parasitaemia when compared with both low and moderate parasitaemia. The differences were statistically significant ($P \leq 0.05$). This agrees with the results obtained by [33] who reported that hyperparasitaemia is associated with higher serum bilirubin level along with increased incidence of complications such as anaemia, haemoglobinuria leading to black water fever and acute renal failure. The role of liver injury or hepatocellular damage in patients has been proposed by many workers [34,35]. However in this study about 38 (5.5%) children with severe malaria (percentage parasitized red cells >5%) had conjugated hyperbilirubinaemia and this probably suggests hepatopathy. In an earlier work reported by [36], it was shown that the incidence of malaria hepatopathy in children

with severe malaria was 8%, while [37] reported 32%. This study suggests that there was an element of hepatic dysfunction characterized by rise in serum conjugated bilirubin especially in the absence of hepatotoxic drugs exposure. Centribular liver damage is one of the factors suggested to have been involved in hepatic dysfunction in acute malaria infection leading to hyperbilirubinaemia which seemed to be a direct consequence of the impaired drainage capacity of the liver, [38] because of suppression of bilirubin excretion due to the effect of parasitaemia on the hepatocyte or endotoxemia or metabolic acidosis.

It was observed that aspartate aminotransaminase (AST) was elevated in the malaria infected subjects when compared with the control group and the difference was statistically significant ($P \leq 0.05$). This result agrees with the earlier findings of [39,21]. Similarly, there was higher level of AST in high parasitaemia more than in both low and moderate parasitaemia. The differences were statistically significant ($P < 0.05$). This result agrees with earlier findings of [39].

Lower levels of albumin were observed in the malaria infected children when compared with

the non infected children. This observation agrees with the work of [40,41,42], who reported that subjects infected with malaria, had their serum albumin levels dropped by 15%. However, Asian [42] suggested that the decrease in albumin level may reflect the acute phase reaction and may help in determining the prognosis on admission. It was not then a surprise to observe higher levels in albumin at lower parasitaemic subjects as against those with moderate parasitaemia which was statistically significant ($P \geq 0.05$). The higher albumin levels may have accounted for the non severity of the fever and other symptoms of malaria at that level. This result agrees with the report of [41]; which stated that at high parasitaemia, there was a decrease in albumin level when compared with low and moderate parasitaemia but their's was not statistically significant ($P \geq 0.05$). The effects of albumin on these complications may be one of the mechanisms by which albumin infusion achieves faster recovery from hypovolemia due to malaria than synthetic colloidal infusions.. Therefore, it is suggested that malaria patients, should be administered albumin infusion specially children with severe malaria because when resuscitated with albumin infusion results in a lower mortality than when with other synthetic colloidal infusions. This may be attributed to the colloidal properties of albumin; possible specific neuroprotective effect, its improved microvascular perfusion in malaria through its rheological effects, volume expansion and influencing fluid shifts across the endothelium.

5. CONCLUSION

The significant reduction in levels of serum albumin and increase in levels of TB, UNB, CB and AST which are liver function parameters indicate that high malaria parasitaemia has some significant adverse effect on the integrity and functions of the liver among children which may lead to mortality if neglected.

CONSENT

As per international standard or university standard, patient's written consent has been collected and preserved by the authors.

ETHICAL APPROVAL

As per international standard or university standard, written approval of Ethics committee

has been collected and preserved by the authors.

ACKNOWLEDGEMENT

The authors wish to gratefully acknowledge the staff of Palmars, Omega and Braithwite Memorial Specialist Hospitals for assistance in sample collection. We appreciate also moral support from colleagues.

COMPETING INTERESTS

The authors declare that there were no conflict of interest or bias at the course of this research. I certify that all authors have participated sufficiently in the conception and design of this work. While it was funded by the researchers, the work was done in Rivers State University, Port Harcourt, Nigeria.

REFERENCES

1. Samba EN. The burden of malaria in Africa. *African Health*. 1997; 19:17 – 18.
2. Rosenthal P. Defining and defeating the intolerable burden of malaria. *Progress and prospective*. *American Journal of Tropical Medicine and Hygiene*. 2007;77:1 – 327.
3. Ogun SA. Management of malaria. *The Nigeria Medical Practitioner*. 2006;49(5): 112-115.
4. Hartman TK, Rogerson SJ, Fisher PR. The impact of maternal malaria on newborns. *Annals of Tropical Paediatrics*. 2010;30 (4): 271-282.
5. World Health Organization. *World Malaria Report*; 2010. Available:<http://www.who.int>
6. Midega JT, Mbogo CM, Mwambi H, Wilson MD, Ojwang G, Wangang JM, Nzovu JG, Githune J, Yan G, Beler JC. Estimating dispersal and survival of *Anopheles gambiae* and *Anopheles funestus* along the Kenyan coast by using mark-release-recapture method. *Journal of Medical Entomology*. 2007;44(6):923-929.
7. Cox-Singh J, Davis TM, Lee KS, Samsul SS, Matusop A, Ratnam S, Rahman HA, Conway DJ, Singh B. *Plasmodium knowlesi* in humans is widely distributed and potentially life threatening. *Clinical Infectious Disease*. 2008;46(2):165-171.
8. Annan Z, Durand P, Ayala FJ, Mathau C, Awono-Ambene P, Sina J. Population genetic structure of *Plasmodium*

- falciparum* in the two main African vectors, *Anopheles gambiae* and *Anopheles funestus*. Proceedings of the National Academy of Science. 2007;104(19):7987-7992.
9. Angella AF, Salgueiro P, Gil LHS, Vicente LJ, Pinto J, Ribolla EMP. Seasonal genetic partitioning in the neotropical malaria vector, *A. darlingi*. *Malaria Journal*. 2014; 13:203.
 10. Wokem GN, Nwachukwu BC, Jackreece, OPJ. Studies on *Plasmodium* Parasitaemia in Ataba-Adoni LG. A. of Niger Delta Region, Rivers State, Nigeria. *Niger Delta Biologia*. 2006;6(1):10-13.
 11. Wokem GN, Okafor RA, Nwachukwu BC. Some haematological profiles of children with malaria parasitaemia in Port Harcourt, Nigeria. *Nigerian Journal of Parasitology*. 2008;29(2):92-97.
 12. Olasehinde GI, Adekeye BT, Ajay AA, Taiwo SO, Adeyeba OA. Management of *falciparum* malaria among infants and children in Ota, Ogun state, Southwest, Nigeria. *African Journal of Clinical and Experimental Microbiology*. 2010;11(3): 689.
 13. Kalu MK, Obasi AN, Nduka FO, Otuchristian G. A comparative study of the prevalence of malaria in Aba and Umuahia. *Research Journal of Parasitology*. 2012;7:17-24.
 14. World Health Organization. World Malaria Report; 2014. Available: <http://www.who.int>
 15. World Health Organization. Malaria report 2017– Malaria World; 2017. Available: <https://malariaworld.org>
 16. Ogbadoyi E, Tsado RD. Renal and hepatic dysfunction in malaria patients in Minna. *Online Journal of Health Allied Sciences*. 2009;8(3):8.
 17. Rijam A, Tor-Agbidye S. The challenges of malarial infection in Nigeria. *The Nigerian Clinical Review Journal*. 2011;89(3):6-34.
 18. Rajesh B, Purmina L, Gehlot RS. Liver involvement in *falciparum* malaria. A histopathological analysis. *Indian Academy of Clinical Medicine*. 2003;4(1):34-38.
 19. Bhalla A, Suri V, Singh V. Malaria hepatopathy. *Journal of Post Graduate Medicine*. 2006;52(4):315-320.
 20. Pironi C, Quirie J, Martin E, Pnesta P, Horacio A, Lee DW. Animal pigment bilirubin discovered in plants. *Journal of the American Chemical Society*. 2009; 131(8):2830.
 21. Onyesom I, Onyemakonor R. Level of parasitaemia and changes in some liver enzymes among malaria infected patients in Edo-Delta Region of Nigeria. *Current Research Journal of Biological Science*. 2011;3(2): 78-81
 22. Maduka C, Neboh I, Emeka E, Eyisi B. Effect of malaria parasitaemia on liver enzyme tests. *International Journal of Tropical Medicine*. 2008;3(3):9-14.
 23. Burtis C, Ashwod E, Border B. *Fundamentals of clinical chemistry: Liver function*. Philadelphia, Saunders Elsevier; 2001.
 24. Araoye MO. *Research methodology with statistics for health and social science: Subjects selection*. Ilorin: Nathadex; 2003.
 25. Mally HT, Evelyn KA. The determination of bilirubin with the photoelectric colorimeter. *Journal of Biological Chemistry*. 1937; 112(2):481– 491.
 26. Martinek R. Improved micro-method for determination of serum bilirubin. *Clinical Chemistry Acta*. 1996;13:61-170.
 27. Reitman, S, Frankel, S. (1957). Colorimetric method for the determination of serum glutamicoxaloacetic and glutamic pyruvic transaminase. *American Journal of Clinical Pathology*, 28, 56-63.
 28. Grant, G.H. & Kachmer, J.S. (1987). *Amino acids and proteins: Fundamentals of Clinical Chemistry*. Philadelphia, USA, W.B Saunders.
 29. Cheesbrough M. *District laboratory practice in tropical countries: Transmission and life cycle of malaria parasite*. United Kingdom: Cambridge University Press; 2006.
 30. White NJ, HO M. The pathophysiology of malaria. *Advanced Parasitology*. 1992;31: 84-167.
 31. Kausar MW, Moeed K, Asit N, Rizwi F, Raza S. Correlation of bilirubin with liver enzymes in patients of *falciparum* malaria. *International Journal of Pathology*. 2010;8 (2):63-67.
 32. Mehta SR, Naidur G, Chander V, Singh IP, Joshi S, Ahuja RC. *Falciparum malaria. Present day problem. An experience with 44424 cases*. *Journal of Indian Physicians Association*. 1989;37:264-266.
 33. Kochar DK, Agarwal P, Kochar SK, Jain K, Rawat N, Pokharana R. Hepatocyte dysfunction and hepatic encephalopathy in *P. falciparum* malaria. *Quarterly Journal of Medicine*. 2006;96(7):505-512.

34. Chawla LS, Sidhu G, Sabharwal BD, Bhatia KL, Sood A. Jaundice in *P. falciparum*. Journal of Indian Physician Association. 1989;43:206-208.
35. Kochar DK, Singh P, Agarwal P, Kochar SK, Pokharna R, Sareen PK. Malaria hepatitis. Journal of Indian Physicians Association. 2003;51:1069-1072.
36. Bag S, Samal GC, Deep N, Patra UC, Nayak M, Mehar LK. Complicated falciparum malaria. India Paediatrics. 1994;31:821-825.
37. Adekunle AS, Adekunle OC, Egbewale BE. Serum status of selected biochemical parameter in malaria. An animal model. Biomedical Research. 2007;18(2):109-111
38. Bhave SY, Joshi SV, Warad V, Dhar HL. Hepatic and renal dysfunction in childhood malaria. Bombay Hospital Journal. 2005; 45:79-84.
39. Golden M. Transport proteins as indices of protein status. American Journal of Clinical Nutrition. 1982;35:1159-1165.
40. Ogbodo SO, Okeke AC, Osu HA, Shu E.N. Chukwurah EF. Nutritional status of parasitemic children from malaria endemic rural communities in eastern Nigeria. Current Paediatric Research. 2010;14(2): 131-135.
41. Ahsan T, Ahmad N, Shaheer A, Mahmood T, Ali H, Farooq MU, Bkaht SF. Jaundice in falciparum malaria. Changing trends in presentation, need for awareness. Journal of Pakistan Medical Association. 2008;58: 616.
42. Akech S, Gwe S, Idro R, Fegan G, Eziefula AC, Charles RJ. Volume expansion with albumin compared to gelofusine in children with severe malaria. Result of a controlled trial. Public Library of Science Clinical Trials. 2006;1(5):21.

© 2018 Wokem et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

*The peer review history for this paper can be accessed here:
<http://www.sciencedomain.org/review-history/25641>*