



Diversity, Life Forms, Chorology and Uses of Spontaneous Medicinal Plants in Niamey and Tillabéri Regions, Niger Republic

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Authors' contributions

This work was carried out in collaboration among all authors. Author AJM designed the study, performed the statistical analysis, wrote the protocol and wrote the first draft of the manuscript. Authors SD and MMI managed the analyses of the study. Author SM managed the literature searches. All authors read and approved the final manuscript.

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ABSTRACT

Plants are the pharmacy of people in rural and urban areas in West African. However, few studies assessed the ecology and diversity of medicinal plants mostly rural and urban areas. This study assessed the taxonomic composition, life form, chorology and uses of the different organs of the spontaneous medicinal plants in Niamey and Tillabéri regions in Niger. The questionnaire survey consisted of individual interview on 168 traditional healers in two regions. A total of 181 species belonging to 60 families were recorded in Niamey and Tillabéri regions where Fabaceae (19

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species) was the dominant family in two regions. The woody species presented 56.59% of the total recorded flora while the herbaceous species were 43.41%. The biological spectrum showed that phanerophytes (57%) were the dominant followed by therophytes (24%) in both regions. This indicates the use of trees, shrubs and annual plants in the traditional medicines in two regions. It also indicates about the tropical habitat. Chorological analysis results revealed that most of the taxa are Sudano-Zambezian (38%) at African scale. Whereas at global level African species (62%) were the dominant chorotype. Leaves (36%) were the most used parts followed by the bark (20%). These different proportions of uses show that the pharmacopoeia of the study area is based on African savannah trees and shrubs. Given the diversity of plant used traditional medicines recorded in two regions, the study recommends successful integration of the use of medicinal plants into a public health framework for biodiversity conservation in both regions. It is important to strengthen conservation strategies to protect this heritage.

Keywords: Ethnobotany; medicinal plants; life forms; chorology; Sahel; biodiversity; Niger.

1. INTRODUCTION

Medicinal plants offer the considerable socio-economic potential to people in developing countries. They are even more important because they are used by about 80% of populations for primary health care [1,2]. They include all plants that contain one or more substances that can be used for therapeutic purposes or that are precursors to the synthesis of useful drugs [3]. Better still, these plant genetic resources play a cultural role for populations, sometimes with religious functions [4].

In addition, the recurrence of rainfall deficits in recent decades has led to a decline in food production mainly based on rainfed crops [5]. Natural ecosystems are therefore subject to increased exploitation by a population seeking dietary supplements and sources of income, but also from strong pastoral pressure [6]. There is an increasing reduction in plant resources and some species are threatened with extinction worldwide because of climate change and pressure from human activities [7,8,9,10]. This is the case in Niger, where plant resources subject to recurrent droughts and climate change impacts, are undergoing a strong and growing anthropogenic pressure in the face of an ever-increasing population with a population growth rate of 3.83%. This in turn reduces plant biodiversity in general and affects that of medicinal plants in particular [11]. An ethnobotanical survey conducted in the regions of Niamey and Tillabéri showed that they contain nearly 2/3 of the country's forest resources showed that many plants are used in remedies [12]. The fundamental question that arises is how can we reconcile the sustainable exploitation of medicinal spontaneous species and their conservation in a context of socio-economic transformation, strongly linked to climate

variability? To answer this question, it is imperative to know the potentialities still available in our forest formations. This knowledge can serve as a baseline situation for the implementation of specific actions aimed at the strategic conservation of these plant species. This study is part of this approach. It aims to assess the ecological diversity of medicinal plants in the Niamey and Tillabéri regions. It is a question of determining their life forms (biological types), their world and African chorologies, as well as the organs harvested for the purposes of the pharmacopoeia.

2. MATERIALS AND METHODS

Ethnobotanical data were collected using purposive sampling based on the sex, age, their origin of knowledge (heritage from the family or out of the family), duration of the exercise, parallel professions, farmers, traditional healers, and other occupations in both regions. The questionnaire survey consisted of individual interview on 168 traditional healers in two regions. The ethnobotanical questionnaires touched the name of the species, the different parts of plants harvested used for medicinal purpose. For biological spectrum and chorological data were collected from the list of the species recorded during the questionnaire and secondary data from published papers. They have been determined for each taxon the life forms, the phytogeographical types in Africa and in the world. This description was developed using scientific documentation [13,14]. For the life forms, we used those classified by Raunkier's and adapted by [15,16] as follow:

2.1 Life Forms

The establishment of the biological spectrum was based on Raunkier's classification adapted by

[15,16].The rate of each of the biological types is evaluated at each station that reflects the conditions of the environment. The definition is based on the position of the buds on the stem and the mechanisms allowing the plants to cross the bad season. Thus, we distinguish according to this author:

1. **Phanerophytes (P):** they are woody, perennial plants whose buds are located more than 50 cm above the ground; they are classified into several types:

- nanophanerophytes (np) that are between 50 cm and 2 m height;
- microphanerophytes (mp) 2 to 8 m height;
- mesophanerophytes (mP) from 8 to 30 m tall;
- nanophanerophytic creepers and microphanerophytic creepers which are lianas;

2. **Chamephytes (CH):** These are woody or suffrutescent species whose buds are located at a maximum of 50 cm from the ground.

3. Hemicryptophytes are plants whose buds are at ground level.

4. **Geophytes:** These are plants whose perennial organs are buried in the soil, organs from which grow herbaceous overhead stems during the wet season that dry out and fall in the dry season. The organs will remain in the ground to give other stems in the next wet season. Geophytes are distinguished by the nature of the perennial organ:

- rhizomatous geophytes (Gr)
- bulbous geophytes (Gb)
- Tuberculous geophytes (Gt)

5. **Therophytes (T):** These annual plants complete their cycle of development during the wet season to live in the seed state during the dry season.

6. **The lianas:** Liane-microphanerophyte (Lmp) liane-chaméphyte (LCH).

2.2 Phytogeographical Spectrum

The phytogeographical distribution of species based on their chorological affinities has been described at the African and global levels. In African phytogeographical zones, we have the affinities:

1. Guino-congolian (GC);
2. Guino-congolian-Sudano-Zambezi (GC-SZ);
3. Sudano-Zambezi (SZ);
4. Sudano-zambezi-Saharo-sindian (SZ-Sah.S);
5. Saharo-sindian (Sah.S);
6. Guineo-Congolian-Sudano-Zambezi-Saharo-sindian (GC-SZ-Sah.S);
7. Saharo-sindian- mediterranean (Sah.S-Med);
8. Sudano-Zambezi-Saharo-Sindian-Mediterranean (SZ-Sah.S-Med).

In the World phytogeographical zones, we have the affinities:

1. African (A);
2. Pantropical (Pt) existing throughout the tropical zone in Africa, Asia and America;
3. Palaeotropical (Pal) species common to Asia, tropical Africa and some islands of the Indian;
4. Afro-Malagasy (AM) common to Africa in Madagascar to the Comoros Islands, Mascaraignes Islands and Seychelles;
5. Cosmopolitan (Co) species whose range covers the entire world;
6. Afro-Neotropical (An), common to Africa and tropical America.

2.3 Data Analysis

We analyzed the data using descriptive statistics in Excel to determine the percentage of each family based on the species richness, the percentage of each life form, chorotypes and the proportion of the organs used the pharmacopoeia in the two regions.

3. RESULTS AND DISCUSSION

3.1 Diversity of Medicinal Plants

The study recorded 181 of spontaneous medicinal species belonging to 60 families; the most represented were the *Fabaceae* (19 species with 10.50%), *Asclepiadaceae*, *Caesalpiniaceae*, *Combretaceae* and *Mimosaceae* (9 species each with 4.97%), *Capparaceae*, *Euphorbiaceae*, *Poaceae* and *Rubiaceae* (8 species each, with 4.42%) from 181 species. A total of 29 families were monospecific as shown in Table 1.

The dominance of *Fabaceae* reported in agroforestry systems in Niger which dominance

was due to the socio-economic of the family [17,18]. For instance, [18] reported that the dominance of *Fabaceae* in agroforestry systems was due to its socio-economic importance such as one of the uses is medicinal utilization. Such utilization may explained the dominance of this family in our study area. Such dominance may be explained according to the findings of Molares et al. [19] who reported that the *Fabaceae* species are of great medicinal importance in indigenous and urban communities. This dominance could be due to the adaptability of the *Fabaceae* species to support large ecological amplitude in terms of climate and soils as reported by Clarice et al. [20]. This dominance is an indication of diversity of a given environment [21]. It is also of great importance to vitality of a given ecosystem as *Fabaceae* species are nitrogen-fixing plants. Added to that, *these families* have been found in abundance in the forest galleries of the Biosphere Reserve of Ouedraogo [22] conducted a study from *Hippopotamus* stream in Burkina Faso witch reflecting a similar results on the flora. Thus, this author has counted about 82 medicinal plant species, belonging to 35 botanical families of which six were found most frequently cited *Combretaceae* (15%), *Caesalpinaceae* (10%), *Mimosaceae* (7%), *Rubiaceae* (6%), *Moraceae* (5%) and *Fabaceae* (5%). However, our results contradict some findings. For instance, in Cameroon, Mpondo (2017) recorded 90 medicinal plant species divided into 47 families and 83 genera, with a clear dominance of the

Euphorbiaceae family (11.11%), followed by *Fabaceae* and *Malvaceae* (5.55% both) and *Rubiaceae* (4.44%). Bouziane [23] reported from Algeria, the dominance of *Lamiaceae* (18 genera), *Apiaceae* (14 genera), *Asteraceae* (10 genera), *Rosaceae* (7 genera) among 134 species belonging to 57 families were recorded. According to Tahri et al. [24], in an ethnobotanical survey conducted in Settat (Morocco), 90 species belonging to 44 families, with the most represented being *Lamiaceae* (17.58%), *Apiaceae* (9.89%), *Asteraceae* (6.59%), *Fabaceae* (5.49%), *Myrtaceae* and *Poaceae* (4.39% each) were recorded, *Lamiaceae* predominate, followed by the *Apiaceae* family, *Asteraceae* with 24.61%, 7.68% and 6.14% respectively listed plants [25].

3.2 Ecology of Medicinal Plants

3.2.1 Biological spectrum

The medicinal flora of the Niamey and Tillaberi regions includes 14 biological types. The spectrum is dominated by phanerophytes (more than 50% of species) followed by therophytes (24%) and chaméphytes (6%). Among the phanerophytes, the microphanerophytes are the most dominant (34% of the species) then the mesophanerophytes and the nanophanerophytes (9% each) as indicated (Fig. 1). The dominant morphological types were woody species (56.59%) while herbaceous plants presented 43.41% of the total flora.

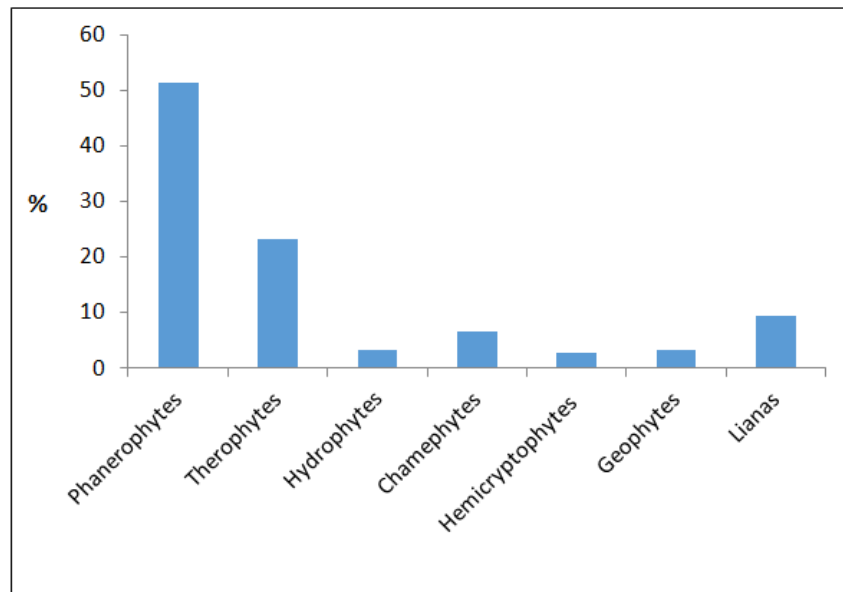


Fig. 1. Biological spectrum of the medicinal plant species recorded in study areas

Table 1. Species number and percentage per family

Families	NB species	%
Fabaceae	19	10.50
Asclepiadaceae	9	4.97
Caesalpiniaceae	9	4.97
Combretaceae	9	4.97
Mimosaceae	9	4.97
Capparaceae	8	4.42
Euphorbiaceae	8	4.42
Poaceae	8	4.42
Rubiaceae	8	4.42
Asteraceae	5	2.76
Cucurbitaceae	5	2.76
Acanthaceae	4	2.21
Anacardiaceae	4	2.21
Convolvulaceae	4	2.21
Moraceae	4	2.21
Bombacaceae	3	1.66
Meliaceae	3	1.66
Pedaliaceae	3	1.66
Scrophulariaceae	3	1.66
Solanaceae	3	1.66
Tiliaceae	3	1.66
Vitaceae	3	1.66
Amaranthaceae	2	1.10
Arecaceae	2	1.10
Bignoniaceae	2	1.10
Burseraceae	2	1.10
Elatinaceae	2	1.10
Lamiaceae	2	1.10
Malvaceae	2	1.10
Rhamnaceae	2	1.10
Sterculiaceae	2	1.10
Agavaceae	1	0.55
Aizoaceae	1	0.55
Amaryllidaceae	1	0.55
Annonaceae	1	0.55
Araceae	1	0.55
Asparagaceae	1	0.55
Balanitaceae	1	0.55
Boraginaceae	1	0.55
Salicaceae	1	0.55
Chrysobalanaceae	1	0.55
Cochlospermaceae	1	0.55
Commelinaceae	1	0.55
Cyperaceae	1	0.55
Ebenaceae	1	0.55
Flacourtiaceae	1	0.55
Hyacinthaceae	1	0.55
Hypocrateaceae	1	0.55
Loganiaceae	1	0.55
Loranthaceae	1	0.55
Menispermaceae	1	0.55
Nympheaceae	1	0.55
Olacaceae	1	0.55
Periplocaceae	1	0.55

Families	NB species	%
Polygalaceae	1	0.55
Pontederiaceae	1	0.55
Salvadoraceae	1	0.55
Sapindaceae	1	0.55
Sapotaceae	1	0.55
Tribulaceae	1	0.55
Total	181	100

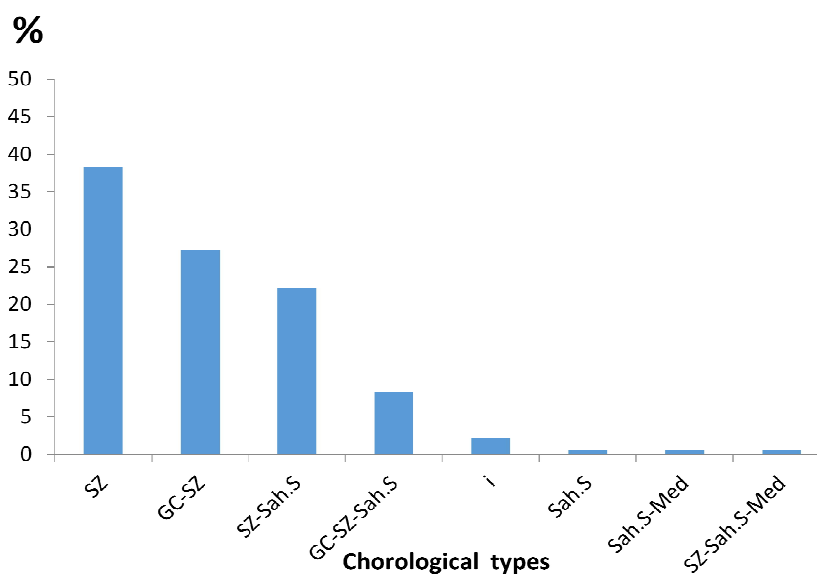


Fig. 2. Chorological types spectrum at African level of medicinal species in the study areas
 Legend: 1. Guino-congolian (GC); 2. Guino-congolian-Sudano-Zambeian (GC-SZ); 3. Sudano-Zambeian (SZ);
 4. Sudano-zambeian-Saharo-sindian (SZ-Sah.S); 5. Saharo-sindian (Sah.S); 6. Guineo-Congolian-Sudano-Zambeian-Saharo-sindian (GC-SZ-Sah.S); 7. Saharo-sindian-mediterranean (Sah.S-Med); 8. Sudano-Zambeian-Saharo-Sindian-Mediterranean (SZ-Sah.S-Med)

The dominance of phanerophytes in the medicinal flora of the two regions could be due the permanence of those plants throughout year. One of the practical of implication of this dominance of the some part of the phanerophytes can be harvested for medicinal purpose at any season during the year. The phanerophytes in this study consists of the dominance of microphanerophytes, plants with 2 to 8 m height that are composed of trees and shrubs. This dominance suggests the crucial role that woody species play in traditional medicines as reported by Abate et al. [26]. However, the dominance of the therophytes indicates the use of annual plants in traditional medicine. In our study, the dominance of phanerophytes and therophytes indicates about the tropical environment of the two regions. For instance, there is also a strong presence of therophytes, well adapted to the ecosystems of the study area (North-Sudan) marked by some aspect of aridity.

An optimal annual stock of therophyte medicinal plants can easily be built up at a specific time, the end of the rainy season. In addition to that, Mpondo et al. [27] found out 11 biological types with megaphanerophytes dominance estimated at 12.22%. Furthermore, according to Adomou et al. [28] in their study on medicinal plants sold in the market of Abomey-Calavi (Benin), 68% of species are phanerophytes. N'Guessan et al. [29] made the similar observations on emmenagogue plants used in traditional medicine, also by Gnagne et al. [30] on plants used in the treatment of diabetes, and even Sylla et al. [31], with respectively 90.90% and 85.18%. In fact, phanerophytes represent 80 to 90% of biological types in tropical and equatorial zones [32,33], which would explain these different results. The more importance biological types, according to the inventory of Bouziane [23] are hemicryptophytes and phanerophytes.

The woody plants are the most abundant morphological type in our study (103 woody species). These biological and morphological characteristics make it possible to ensure a permanent availability of the raw material used in the pharmacopoeia. These results corroborate those of Dibong et al. [34] which reported that woody plants represented (60%) of the study area, as well as Sylla et al. [31] (81.48% of woody species), Kouchadé et al. [35] (53.68% woody plants). Added to that are 33.33% trees among the identified plants [27]. Ambé et al. [32] stated that the plants used are mainly trees and shrubs (74.6%). Several other authors have noted the importance of ligneous plants in medicinal flora [36,37].

3.2.2 Phytogeographical spectrum

At African level, the chorological spectrum analysis of medicinal plants in our study revealed 9 African chorotypes mainly Sudano-Zambezian (38%), Guineo-congolian-Sudano-Zambezian (27%), Sudano-Zambezian-Saharo-Sindian (22%), Guineo-Congolian-Sudano-Zambezian-Saharo-Sindian (8%) species for the spectrum (Fig. 2).

The chorology analysis results at global level on the medicinal plants in the study area showed

that African species come first, followed by paleotropical and then pantropical species (Fig. 3). The results give clearly preponderance of African species then of widely distributed species: paleotropical and pantropical (29%) followed by afro-tropical (10%) [28]. Afro-tropical species (34.44%) are the most cited [27]. A high proportion of widely distributed species may be a degradation index [38]. This describes the chorological affinities of the medicinal species in the study area, but also proves that the pharmacopoeia bequeathed to us is based on African plants well known to our predecessors for having been in contact and observed this heritage for a long time. The medicinal flora studied must be described essentially from an African way. Thus, among the 9 phytogeographical types characterizing the African chorology of our medicinal plants (three phytogeographical types), the Sudano-Zambezian (38%), Guineo-Congolian-Sudano-Zambezian (27%), and Sudano-Zambezian-Saharo-Sindian (22%) are the most encountered. The geographical location of our study area, distributed between the Sudan and Saharan regions explains the strong presence of the binding species (GC-SZ, SZ-Sah. S.). The high proportion of Sudano-Zambezian species in the medicinal flora shows that it corresponds to a Sudanian pharmacopoeia. However, despite the

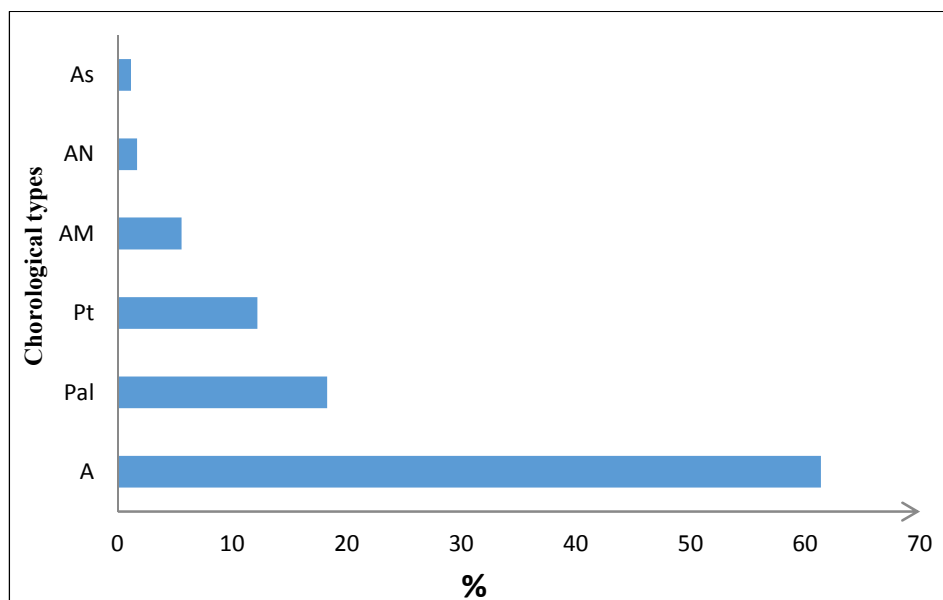


Fig. 3. Chorological types spectrum at World level of medicinal species in the study areas
 Legend: 1. African (A); 2. Pantropical (Pt) existing throughout the tropical zone in Africa, Asia and America; 3. Palaeotropical (Pal) species common to Asia, tropical Africa and some islands of the Indian; 4. Afro-Malagasy (AM) common to Africa in Madagascar to the Comoros Islands, Mascaraignes Islands and Seychelles; 5. Cosmopolitan (Co) species whose range covers the entire world; 6. Afro-Neotropical (An), common to Africa and tropical America

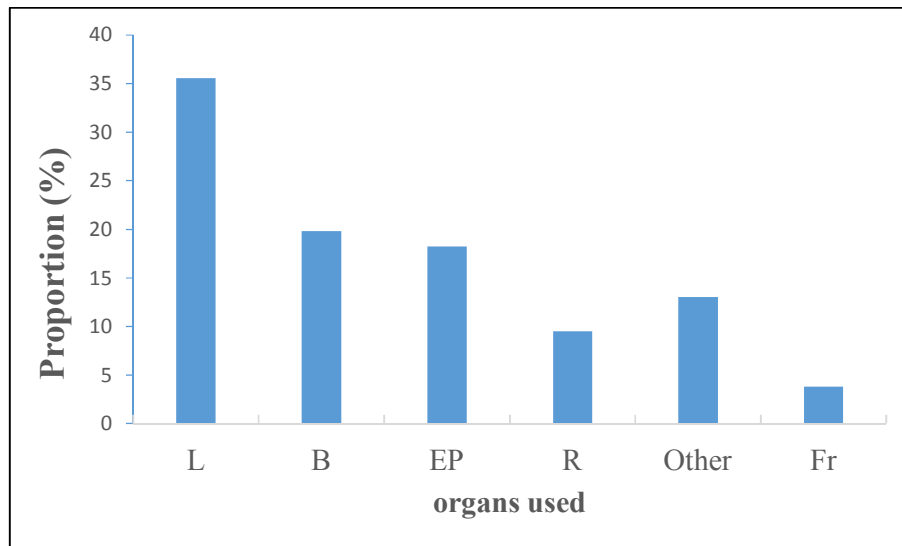


Fig. 4. Proportion of different plant parts used in pharmacopoeia

Legend: L = Leaves, B = barks, EP = Entire plant, R = root, Fr = flower, Other = organ associations.

shifting of isohyets towards the south for several decades [11,39], the local cultures built for a long time are maintained mainly around the species of more or less humid zones (SZ and GC -SZ).

According to Sylla et al. [31], the Guineo-Congolian-Sudano-Zambeian species are the most important (48.15%), a similar result to those obtained in other studies [32]: 43%, [30]. Further, Betti and Mebere [40] reports significant dominance of Guinean species (67.54%), as well as [28] who inventoried 29% of Guinean people. For N'Guessan et al. [29], also the medicinal plants studied are Guineo-Congolian (54.54%) and Sudano-Zambeian (36.36%). While Bouziane [23] affirms the abundance of the Mediterranean elements. The plants introduced represent 18% in the study of Gnagneet al. [30] ; 9.09% according to N'Guessan et al. [29], unlike our work where they are only 2%. The influence of foreign cultures is therefore not remarkable in the flora studied.

3.3 Uses of Plant Organs in Pharmacopoeia in Niamey and Tillabéri Regions

3.3.1 Harvested plant parts

Our analysis showed that each parts of recorded plant species is used as medicine. Nevertheless, the most used parts are the leaves (36%), the bark (20%), or the entire plant (18%) (Fig. 4). Organs are rarely used in mixture with each other or with exudates (gum, latex).

Leaves and bark are the preferred parts, followed by the entire plant and root. Leaves are the part of the plant commonly used with 38.46% [29], and 39.33% according to El Hilah et al. [25]. In 41% of cases, the listed medicinal plants were solicited for their leaves [41]. For Sylla et al. [31] leaves are the most used organs (68.89%) in the treatment of malaria in the district of Abidjan. Bouziane [23] reports that leaves are the most used part (46.30%), followed by fruits and seeds (26.17% each) and flowers (21.13%). [42,36] have also asserted the primacy of leaves in the pharmacopoeia compared to other plant organs.

Debarking is very important for the species *Pteleopsis suberosa* (Fabaceae) in which a whole branch is completely stripped of its cortical part. Woody species are most often asked for their bark or their roots [27]. The choice of these organs is explained by their ease in being harvested and their availability in all seasons [28]. The ease and speed of harvest can explain the high frequency of leaf use [43].

The organs are chosen according to the specialty [44]. Ouedraogo and Abate et al. [22,36] has reported the preferential use of leaves for medicinal preparations intended for children, and roots for the care of adults This practice may have a role in the conservation of certain species. Indeed, if the removal of the leaves of a tree to a certain extent (up to 50%) does not affect its existence [41], the exploitation of the bark or the root can cause a great risk for its survival or for its ecosystem [45,46].

In addition, the harvest is anarchic, regardless of the amount to be taken, or a suitable technique as noted by [27]. The herbaceous plants are completely unearthed; the only alternative for the survival of these species is the harvest after production and release of diaspore. Sellers of medicinal plants collect species that cause the early death of trees, shrubs, herbs and lianas [38]. The removal of organs or whole plants for the purposes of the pharmacopoeia is a common practice that is an important factor of stress in plants especially for those who are employed in various fields [47] or geophytes [40]. The debarkation of the trunks and branches of some species could explain their important mortality in the botanical reserve of Noflaye (*Khaya senegalensis*, *Prosopis africana*). These species are endangered in the reserve [48]. The removal of the root affects the vegetative aspect as well as the physiology of the plant [49].

Regarding the selection of harvested organs, current knowledge allows us to analyze medicinal plants and often to understand the use recommended by our ancestors [50]. Empirical at the beginning, this use can have scientific bases. Indeed, the leaves, the bark and the roots are generally rich in secondary metabolites responsible for the biological properties of the plants [51,52,53,54].

All the 15 tree species protected under the law establishing the forest regime in Niger belong to the medicinal flora studied. Several authors have reported the use of these plants in various fields (food, craft, real estate ...). Among these highly sought-after species with special status, 4 namely *Sclerocarya birrea*, *Acacia nilotica*, *Ziziphus mauritiana*, *Khaya senegalensis* are among the most commonly used species in traditional medicine [55].

On the study carried out in the W Biosphere Reserve of Niger, several woody species that have become rare or disappeared, much in demand for the traditional pharmacopoeia, were cited by the interviewees. These plants 24 species namely *Abrus precatorius*, *Albizia chevalieri*, *Annona senegalensis*, *Anogeissus leiocarpa*, *Bombax costatum*, *Commiphora africana*, *Detarium microcarpum*, *Diospyros mespiliformis*, *Entada africana*, *Ficus ingens*, *Ficus platyphylla*, *Ficus sycomorus*, *Khaya senegalensis*, *Kigelia africana*, *Lannea acida*, *Lannea microcarpa*, *Parkia biglobosa*, *Prosopis africana*, *Pterocarpus erinaceus*, *Securidaca longipedunculata*, *Tamarindus indica*, *Terminalia*

avicennioides, *Ximenia americana* are among our list of spontaneous medicinal plants. The role of natural reserves and national parks in the conservation of medicinal plants is not negligible [56].

In Niger, the context of climate change and ecosystem degradation exacerbated by anthropogenic effects, leading to a loss of biodiversity [12], could compromise the perpetuity of medicinal plant production if adequate measures are taken in favour of sustainable development are not taken.

4. CONCLUSION

The spontaneous and sub spontaneous medicinal flora of the Niamey and Tillaberi regions is rich in 181 species. It consists mainly of African phanerophytes and therophytes, and then widely distributed (paleotropical and pantropical) with regard to world chorology. The phytogeographical distribution in Africa ranks them among the Sudano-Zambezians and the Guineo-Congolian-Sudano-Zambeziian linkage species in particular. The leaves and bark are the most stressed organs. In some cases, the entire plant is used with fatal consequences on the survival of the plant. It is possible to reconcile the sustainable exploitation of spontaneous medicinal species and their conservation in the studied areas. However, it is necessary and urgent to envisage programs of valorisation of these plants, which associate the conservation and the sustainable exploitation of this inheritance through in particular:

- ✓ The identification of the priority species of our pharmacopoeia, especially the plant species with multiple uses, their ecological characteristics by other ethnobotanical surveys;
- ✓ The sensitization of local stakeholders on the good practice of using medicinal plants to reduce the effect of improper picking on plant survival; preferential organ harvesting, the reduction of which does not affect the life of the plants, for example the replacement of the bark, the root, the whole plant by the leaves when their pharmacological properties are comparable;
- ✓ The integration of medicinal plants into urban reforestation programs.

These are all measures that contribute to the socio-economic development of the regions

studied and our country while allowing future generations to reap the same benefits.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Adjanohoun, et al. Medicine and pharmacopoeia, Contribution to ethnobotanical and floristic studies in Niger. ACCT. 1980;251.
2. Jiofack T, Ayissi I, Fokunang C, Guedje N, Kemeuze V. Ethnobotany and phytomedicine of the upper Nyong Valley forest in Cameroon. African Journal of Pharmacy and Pharmacology. 2009;(4): 144-150.
3. Sofowora A. Medicinal plants and traditional medicine of Africa. Ed. Khartala, Paris. 1996:256.
4. Codjia JTC, Assogbajo AE, Ekue MRM. Diversity and local valorization of Benin's food forest plant resources. Cahiers Agricultures. 2003;12(5):321-331.
5. Habou Rabiou, Abdoulaye Diouf, Babou Andre Bationo, Kossi Novigno Segla Structure of natural stands of *Pterocarpus erinaceus* Poir. in the Sudanese field, in Niger and Burkina Faso. Wood and Forests of the Tropics. 2015; 325(3).
6. Sawadogo I. Fodder resources and representations of pastoralists, evolution of pastoral practices in a protected area context: Case of the Kotchari terroir on the outskirts of the W biosphere reserve in Burkina Faso. Thesis, National Museum of Natural History, Paris, France. 2011; 345.
7. Diederichs N, Geldenhuys C, Mitchell D. The first legal harvesters of protected medicinal plants in South Africa. Science in Africa - Africa's First On-Line Science Magazine. Available:<http://www.scienceinafrica.co.zo/2002/November/bark.htm>
8. Cunningham AB, Mbenkum FT. Sustainability of harvesting *Prunus africana* bark in Cameroon: a medicinal plant in International Trade. People and Plants Working Paper No. 2, Division of Ecological Sciences. UNESCO: Paris France; 1993.
9. Oldfield S, Lusty C, MacKinven A. The World List of Threatened Trees. World Conservation Press: Cambridge; 1998
10. Soehartono T, Newton AC. *Aquilaria* II. The impact of gahuru harvesting in Indonesia. Biol. Conserv. 2001;97:29-41.
11. Ozer P. and Ercicum M. Methodology for a better spatio-temporal representation of the rainfall fluctuations observed in Niger since 1905. Drought. 1995;6(1):103-108.
12. Mamadou AJ, Douma S, Mahamane A, Saadou M. Analysis of the state of floristic diversity of plateaus along a north-south gradient in the Niger W Biosphere Reserve. European Scientific Journal. 2017b;14(3). [ISSN: 1857-7881 (Print)] [e - ISSN 1857-7431]
13. Saadou M. Assessment of biological diversity in Niger: constituent elements of plant biodiversity. National Council for the Environment for Sustainable Development SE / CNEDD, Project NER / 97 / G 31 / A / 1 G / 99 "National Strategy and Action Plan - Biological Diversity". 1998;138.
14. Berhaut J. Flora of Senegal. 2nd more complete edition with the humid forests of Casamance. 1967;485.
15. Aké Assi L. Flora of Ivory Coast: systematic catalog, biogeographic and ecological. I. Boissiera. 2001;57:396.
16. Aké Assi L. Flora of Ivory Coast: Systematic, biogeographical and ecological catalog. II. Boissiera. 2002;58: 441.
17. Saadou M. Vegetation drained Nigerian environments east of the Niger River. State Thesis, University of Niamey. 1990;395.
18. Moussa Soulé, Ado Adamou Matalabi, Ibrahima Djibo Bassirou and Saadou Mahamane, systematic composition, life forms and chorology of agroforestry systems of Aguié department, Niger, West Africa. Journal of Applied Life Sciences International. 2017;8(4):1-12. Article JALSI.29138 [ISSN: 2394-1103] Available: www.sciencedomain.org
19. Molares S, Arenas PM, Aguilar A. Etnobotánica urbana de los productos vegetales adelgazantes comercializados en México DF. Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas. 2012;11(5):400-411.
20. Clarice Mudzengi, Shakkie Kativu, Everson Dahwa, Xavier Poshiwa, Chrispen Murungwen. Effects of *Dichrostachys cinerea* (L.) Wight & Arn (Fabaceae) on herbaceous species in a semi-arid

- rangeland in Zimbabwe. *Nature Conservation*. 2014;7:51–60.
21. Raes N, et al. Legume diversity as indicator for botanical diversity on Sundaland, South East Asia, South African Journal of Botany. 2013;89:265-272. Available:<http://dx.doi.org/10.1016/j.sajb.2013.06.004>
 22. Ouedraogo M. The forest galleries of the Biosphere Reserve of the Hippopotamus Mare in Burkina Faso: Characteristics, dynamics and ethnobotany. Thesis 3rd cycle, University of Ouagadougou. 2008; 279.
 23. Bouziane Z. Contribution to the ethnobotanical study of medicinal plants in the Azail region (Tlemcen -Algeria). Master thesis Faculty of Sciences of Nature and Life, Earth and Universe Sciences, University Aboubakr Belkaïd-Tlemcen, Algeria; 2017.
 24. Tahri N, El Basti A, Zidane L, Rochdi A, Douira A. Ethnobotanical study of medicinal plants in the Province of Settât (Morocco). *J. Forest. Fac.* 2012;12(2):192-208.
 25. El Hilah et al. Ethnobotanical study of medicinal plants used in the treatment of dermatological diseases in the central plateau of Morocco. *J. Appl. Biosci.* 2016; 98:9252-9260.
 26. Abate Assefa, Baye Gelaw, Gebeyaw Getnet, Gashaw Yitayew. The effect of incident tuberculosis on immunological response of HIV patients on highly active anti-retroviral therapy at the university of Gondar hospital, northwest Ethiopia: A retrospective follow-up study *BMC Infect Dis.* 2014;14:468. DOI: 10.1186/1471-2334-14-468
 27. Mpondo, et al. Knowledge and Traditional Uses of Medicinal Plants of Upper Nyong Department. *J. Appl. Biosci.* 2017;113: 11229-11245.
 28. Adomou AC, Yedomonhan H, Djossa B, Legba SI, Oumorou M, Akoegninou A .. Ethnobotanical study of medicinal plants sold in the Abomey-Calavi market in Benin. *Int. J. Biol. Chem. Sci.* 2012;6(2): 745-772. DOI: <http://dx.doi.org/10.4314/ijbcs.v6i2.18>
 29. N'Guessan K, Kouadio K, Kouame NF. Emmenagogues plants used in traditional medicine by the Abbey and Krobou peoples of Agboville (Ivory Coast) *Pharm. Med. Trad. Afr.* 2006;137-158.
 30. Gnagne, et al. Ethnobotanical study of medicinal plants used in the treatment of diabetes in the Department of Zouénoula (Côte d'Ivoire). *J. Appl. Biosci.* 2017;113: 11257-11266.
 31. Sylla Y, et al. Ethnobotanical study of plants used against malaria by traditional healers and herbalists in the district of Abidjan (Ivory Coast) *Int. J. Biol. Chem. Sci.* 2018;12(3):1380-1400.
 32. Ambé ASA, Ouattara D, Tiebre MS, Vroh BTA, Zirihi GN, N'Guessan KE. Diversity of medicinal plants used in the traditional treatment of diarrhea in the markets of Abidjan (Ivory Coast). *Journal of Animal & Plant Sciences.* 2015;26(2):4081-4096.
 33. Deleke Koko IK, Djego J, Hounzangbe-Adote MS, Sinsin B. Ethnobotanical study of galactogenic plants and emmenagogues used in riparian lands at the Hunting Zone of Pendjari. *Int. J. Biol. Chem. Sci.* 2009;3(6):1226-1237.
 34. Dibong SD, Mpondo ME, Nigoye A, Kwin MF, Betti JL. Ethnobotany and phytomedicine of medicinal plants from Douala, Cameroon. [Ethnobotany and phytomedicine of medicinal plants sold in Douala markets], *Journal of Applied Biosciences.* 2011;37:2496-2507. [ISSN: 1997-5902] Available: www.biosciences.elewa.org
 35. Kouchadé AS, Adomou AC, Tossou GM, Yédomonhan H, Dassou GH, Akoègninou A. Ethnobotanical study of medicinal plants used in the treatment of childhood diseases and sold in markets in southern Benin. *Journal of Animal & Plant Sciences,* 2016;28(2):4418-4438.
 36. Zerbo P, Millogo-Rasolodimby J, Nacoulma-Ouedraogo OG, Van Damme P. Contribution to the knowledge of medicinal plants used in child care in San country, Burkina Faso. *Int. J. Biol. Chem. Sci.* 2007;1(3):262-274.
 37. Diatta CD, Gueye M, Akpo LE. Medicinal plants used against dermatoses in the Bainnun pharmacopoeia of Djibonker, Senegal. *Journal of Applied Biosciences.* 2013;70:5599-5607.
 38. Adingra OMMA, Kassi JN'D, Yongo OD. Systematic and phytogeographic analysis of the Bamo Classified Forest (Ivory Coast). *Journal of Animal & Plant Sciences.* 2014,23(2):3626-3636. Available: <http://www.m.elewa.org/JAPS> [ISSN: 2071-7024]

39. Fodé M, Amadou OM. Drought and rainfall variations in Niger from 1950 to 1991. *Annals of the UAM*. 2003;8:117-132.
40. Betti J, Mebere S. Contribution to knowledge of non-timber forest products in Kalamaloué National Park, Far North Cameroon: Food plants. *International Journal of Biological and Chemical Biosciences*. 2011;5(1):291-303.
41. Ouattara D. Contribution to the inventory of significant medicinal plants used in the region of Divo (Southern forest of the Ivory Coast) and to the diagnosis of the pepper tree of Guinea: *Xylopiya aethiopica* (Dunal) A. Rich. (Annonaceae). PhD thesis, University of Cocody, Abidjan, 2006;184.
42. Zirihi G. N.,. Contribution to the identification, identification and knowledge of some plant species used in traditional medicine and pharmacopoeia among Bété Department of Issia, Ivory Coast. PhD Thesis, University of Abidjan, F.A.S.T. 1991;150.
43. Bitsindou M. Survey of traditional phytotherapy in Kindamba and Odzala (Congo) and analysis of convergence of use of medicinal plants in Central Africa. Same. Doc (unpublished) Univ. Free from Brussels. 1986;482.
44. Olivier M. Ethnobotanical, pharmacological and phytochemical study of *Cordia dichotoma*, *Hubiscus tiliaceus*, *Savallia solida*. Traditional pharmacopoeia and cutaneous affection in New Caledonia. 2001;23-31.
45. Cunningham AB. Development of the conservation policy on commercially exploited medicinal plant. In *Conservation of Medicinal Plants*, Akerele O, Heywood V, Syngé H (eds). Cambridge University Press, Cambridge. 1991;337-358.
46. Walkers M, Hamilton A. *Plant diversity: A source of vital wealth*. WWF ed., Paris; 1994.
47. Ambouta K. Report on the review of scientific activities in the Niger W Biosphere Reserve and proposal of a research program. 2002;59.
48. Ilboudo JBMH. State and evolutionary trends of the flora and vegetation of the botanical reserve of Noflaye: Elements for a development, thesis of 3rd cycle in Environmental Science, Cheik Anta Diop University, Dakar, Senegal. 1992;107.
49. Yapi AB. Inventory of medicinal plants from the Asteraceae family in the markets of Abobo (Abidjan, Ivory Coast). Master's thesis in botany, Félix Houphouët-Boigny University, Abidjan. 2013;50.
50. Bourrel C. Chymic analysis, biostatistical and antioxidant activities of extracts of selected aromatic plants. Thesis submitted to obtain the degree of Doctor in Biochemistry, National Polytechnic Institute of Toulouse. La France; 1993.
51. Bigendako-Polygenis MJ, Lejoly J. The traditional pharmacopoeia in Burundi. Pesticides and medicines in animal health. *Near. Univ. Namur*. 1990;425-442.
52. N'Guessan K, Kadja B, Zirihi G, Traoré D, Aké-assi L. Phytochemical screening of some Ivorian medicinal plants. *Science & Nature* 2009;6:1-15.
53. Ngono N, Koanga M, Tchinda T, Magnifouet, Motso C, Mballa B. Ethnobotanical survey of some cameroonian plants used for treatment of viral diseases. *African Journal of Plant Science*. 2011;5(1):15-21.
54. Mangambu MJD, Mushagalusa KF, Kadima NJ. Contribution to the phytochemical study of certain antidiabetic medicinal plants of the city of Bukavu and its surroundings (South Kivu, DR Congo). *J. Appl. Biosci*. 2014;75:6211-6220. DOI: <http://dx.doi.org/10.4314/jab.v75i1.7>
55. Hassane H. Répertoire des espèces végétales les plus couramment utilisées en pharmacopée traditionnelle et impact des techniques de prélèvement sur la diversité biologique dans la réserve de Biosphère du W du Niger. Mémoire de DEA Géographie, Milieux et sociétés des espaces arides et semi-arides : Aménagement-Développement. Université Abdou Moumouni de Niamey. 2008. 133 p.
56. Mamadou AJ, Saley K, Boubé M, Rokia S, Saadou M. Ethnobotanical survey of traditional health practitioners from the regions of Niamey and Tillabéri Niger: Data 2012-2017. *European Scientific Journal*. 2017a;13(33):276-304.

Appendix 1. List of families, life forms and chorology of recorded species

Species	Famille	Life forms	Chorology	
			African chorology	World chorology
<i>Abrus precatorius</i> L.	Fabaceae	Lmp	GC-SZ	Pt
<i>Abutilon pannosum</i> (G.Forst.) Schltld.	Malvaceae	np	SZ-Sah.S	Pal
<i>Acacia albida</i> (Del.) A. Chev.	Mimosaceae	mP	SZ-Sah.S	A
<i>Acacia erythrocalyx</i> Brenan.	Mimosaceae	Lmp	GC-SZ	A
<i>Acacia nilotica</i> (L.) Willd.ex Del. Subsp. <i>nilotica</i>	Mimosaceae	mp	SZ	A
<i>Acacia senegal</i> (L.) Willd	Mimosaceae	mp	SZ	Pal
<i>Acacia seyal</i> Del.	Mimosaceae	mp	SZ-Sah.S	A
<i>Acanthospermum hispidum</i> D.C.	Asteraceae	T	iGC-SZ	N
<i>Adansonia digitata</i> L.	Bombacaceae	mP	SZ	A
<i>Albizzia chevalieri</i> Harms.	Mimosaceae	mp	SZ	A
<i>Ampelocissus grantii</i> (Bak.) Planch.	Vitaceae	LGr	SZ	A
<i>Andira inermis</i> (W.Wright.) Kunth ex DC.	Fabaceae	mp	GC-SZ	AN
<i>Annona senegalensis</i> Pers.	Annonaceae	np	SZ	A
<i>Anogeissus leiocarpa</i> (DC.) Guill. et Perr.	Combretaceae	mP	SZ-Sah.S	A
<i>Aristida sieberiana</i> Trin.	Poaceae	H	SZ	A
<i>Asparagus africanus</i> Lam.	Asparagaceae	Gr	SZ	A
<i>Azadirachta indica</i> A. Juss.	Meliaceae	mp	i	Ind
<i>Bacopa crenata</i> (P. Beauv.) Hepper.	Scrophulariaceae	Hy	GC-SZ	AM
<i>Balanites aegyptiaca</i> (L.) Del.	Balanitaceae	mp	SZ-Sah.S	A
<i>Bauhinia rufescens</i> Lam.	Caesalpinaceae	mp	SZ-Sah.S	A
<i>Bergia suffruticosa</i> (Del.) Fenzl.	Elatinaceae	CH	SZ-Sah.S	As
<i>Blepharis linariifolia</i> Pers.	Acanthaceae	T	SZ	Pal
<i>Bombax costatum</i> Pellegr.	Bombacaceae	mP	SZ	A
<i>Borassus aethiopum</i> Mart.	Arecaceae	mP	GC-SZ	A
<i>Boscia angustifolia</i> A. Rich.	Capparaceae	mp	SZ	A
<i>Boscia salicifolia</i> Oliv.	Capparaceae	mp	SZ-Sah.S	A
<i>Boscia senegalensis</i> (Pers.) Lam. Ex Poir.	Capparaceae	mp	SZ-Sah.S	A
<i>Boswellia dalzielii</i> Hutch.	Bursereae	mp	SZ	A
<i>Cadaba farinosa</i> Auct.	Capparaceae	mp	SZ-Sah.S	Pal
<i>Cadaba glandulosa</i> Forsk.	Capparaceae	np	SZ-Sah.S	A
<i>Calotropis procera</i> (Ait) R. Br.	Asclepiadaceae	mp	GC-SZ-Sah.S	Pal
<i>Caralluma adscendens</i> (Roxb.)R. Brown	Asclepiadaceae	CH	SZ	A
<i>Cassia italica</i> (Mill.)F.W. Anders.	Caesalpinaceae	CH	SZ	A
<i>Cassia mimosoides</i> L.	Caesalpinaceae	T	GC-SZ-Sah.S	Pal
<i>Cassia nigricans</i> Vahl.	Caesalpinaceae	T	SZ-Sah.S	Pal
<i>Cassia occidentalis</i> L.	Caesalpinaceae	np	GC-SZ	Pt
<i>Cassia sieberiana</i> DC.	Caesalpinaceae	mp	GC-SZ	A
<i>Ceiba pentandra</i> (L) Gaertn.	Bombacaceae	mP	GC-SZ	Pt

Species	Famille	Life	Chorology	
<i>Celosia trigyna</i> L.	Amaranthaceae	T	GC-SZ	Am
<i>Cenchrus prieuri</i> (Kunth.) Maire.	Poaceae	T	SZ-Sah.S	Pal
<i>Centaurea perrottetii</i> DC.	Asteraceae	CH	SZ	A
<i>Chasmanthera dependens</i> Hochst.	Menispermaceae	Lmp	GC-SZ	A
<i>Chrozophora brocchiana</i> Vis.	Euphorbiaceae	np	SZ-Sah.S	A
<i>Cissus quadrangularis</i> L.	Vitaceae	mp	SZ	Pal
<i>Citullus lanatus</i> (Thumb) Matsumara et Nakai	Cucurbitaceae	LT	SZ-Sah.S	Pal
<i>Cleome gynandra</i> L.	Capparaceae	T	GC-SZ- Sah.S	Pt
<i>Cochlospermum planchonii</i> Hook. f.	Cochlospermaceae	Gr	SZ	A
<i>Combretum aculeatum</i> Vent.	Combretaceae	mp	SZ	A
<i>Combretum glutinosum</i> Perr. ex DC.	Combretaceae	mp	SZ	A
<i>Combretum micranthum</i> G.Don.	Combretaceae	mp	SZ	A
<i>Combretum nigricans</i> var <i>elliottii</i> (Engl.Ex Diels) Aubrev.	Combretaceae	mp	SZ	A
<i>Combretum paniculatum</i> Vent.	Combretaceae	mp	GC-SZ	A
<i>Commelina forskalei</i> Vahl.	Commelinaceae	T	SZ-Sah.S	Pal
<i>Commiphora africana</i> (A. Rich.) Engl.	Burseraceae	mp	SZ-Sah.S	A
<i>Corchorus olitorius</i> L.	Tiliaceae	T	GC-SZ- Sah.S	Pal
<i>Crinum ornatum</i> (Ait.)Burg. Hexandr.	Amaryllidaceae	Gb	GC-SZ	A
<i>Crossopteryx febrifuga</i> (G.Don.) Benth.	Rubiaceae	mp	SZ	A
<i>Crotalaria arenaria</i> Benth.	Fabaceae	T	SZ	A
<i>Crotalaria pallida</i> Ait.	Fabaceae	T	GC-SZ	Pt
<i>Croton zambezicus</i> Müll. Arg.	Euphorbiaceae	mp	GC-SZ	A
<i>Cucumis metuliferus</i> Naud.	Cucurbitaceae	LT	GC-SZ	A
<i>Cucumis prophetarum</i> L.	Cucurbitaceae	LT	SZ-Sah.S	Pal
<i>Cymbopogon giganteus</i> Chiov.	Poaceae	H	GC-SZ	A
<i>Cymbopogon schoenanthus</i> (L) Spreng. subsp. <i>proximus</i> (A. Rich.) Maire et Weiler	Poaceae	H	SZ-Sah.S	A
<i>Detarium microcarpum</i> Gmill. et Perr.	Caesalpiniaceae	mp	SZ	A
<i>Dicoma tomentosa</i> Cass.	Asteraceae	T	SZ	Pal
<i>Diospyros mespiliformis</i> Hochst. Ex A. DC.	Ebenaceae	mp	GC-SZ	A
<i>Dipcadi cowanii</i> (Ridl.) H.Perrier	Hyacinthaceae	Gb	SZ	A
<i>Echinochloa stagnina</i> (Retz.) P. Beauv.	Poaceae	Hy	SZ	Pal
<i>Eichornia crassipes</i> (Mart.) Solms. Laub.	Pontederiaceae	Hy	i	N
<i>Entada africana</i> Guill. et Perr.	Mimosaceae	mp	SZ	A
<i>Eragrostis tremula</i> Steud.	Poaceae	T	GC-SZ- Sah.S	Pal
<i>Euphorbia balsamifera</i> Ait.	Euphorbiaceae	mp	SZ-Sah.S- Med	A
<i>Euphorbia hirta</i> L.	Euphorbiaceae	T	GC-SZ- Sah.S	Pt
<i>Euphorbia sudanica</i> A. Chev.	Euphorbiaceae	np	SZ	A
<i>Evolvulus alsinioides</i> (L.) L.	Convolvulaceae	T	GC-SZ- Sah.S	Pt

Species	Famille	Life	Chorology	
<i>Feretia apodenthera</i> Del.	Rubiaceae	mp	SZ	A
<i>Ficus platyphylla</i> Del.	Moraceae	mP	SZ	A
<i>Ficus polita</i> Vahl.	Moraceae	mp	GC-SZ	A
<i>Ficus sycomorus</i> L. subsp.	Moraceae	mP	SZ-Sah.S	A
<i>Gnaphalocarpa</i> (Miq.) Berg.				
<i>Ficus thonningii</i> Blume.	Moraceae	mP	GC-SZ- Sah.S	A
<i>Fimbristylis hispidula</i> (Vahl.)Kunth. Subsp. <i>hispidula</i>	Cyperaceae	T	GC-SZ	A
<i>Flueggea virosa</i> (Rxb. Ex Willd) Voigt	Euphorbiaceae	mp	GC-SZ	Pal
<i>Gardenia sokotensis</i> Hutch.	Rubiaceae	np	SZ	A
<i>Gardenia ternifolia</i> Schum.et Thonn.	Rubiaceae	np	GC-SZ	A
<i>Glossonema boveanum</i> (Decne) Decne subsp. <i>breveanum</i>	Asclepiadaceae	CH	Sah.S-Med	A
<i>Grewia bicolor</i> Juss.	Tiliaceae	mp	SZ	A
<i>Grewia flavescens</i> Juss.	Tiliaceae	mp	SZ-Sah.S	Pal
<i>Guiera senegalensis</i> J.G. Gmel.	Combretaceae	mp	SZ	A
<i>Gymnema sylvestre</i> (Retz.) Schultes	Asclepiadaceae	Lmp	GC-SZ	Pal
<i>Heliotropium ovalifolium</i> Forssk.	Borraginaceae	CH	SZ	A
<i>Hygophila senegalensis</i> (Nees) T. Anders.	Acanthaceae	Hé	SZ	A
<i>Hygrophila auriculata</i> (Schumach.) Heine	Acanthaceae	Hé	SZ	Pt
<i>Hyphaene thebaica</i> (L.) mart.	Arecaceae	mP	SZ	A
<i>Hyptis spicigera</i> Lam.	Lamiaceae	T	SZ	Pt
<i>Indigofera astragalina</i> DC.	Fabaceae	T	SZ	A
<i>Indigofera berhautiana</i> Gillet.	Fabaceae	T	SZ	A
<i>Indigofera diphylla</i> Vent.	Fabaceae	T	SZ-Sah.S	Pal
<i>Indigofera hirsuta</i> L. var. <i>hita</i>	Fabaceae	T	SZ-Sah.S	Pal
<i>Indigofera leptoclada</i> Harms.	Fabaceae	T	SZ	A
<i>Indigofera tinctoria</i> L.	Fabaceae	np	GC-SZ	Pal
<i>Ipomoea asarifolia</i> (Desr.)Roem.et Schult.	Convolvulaceae	LCH	GC-SZ- Sah.S	Pt
<i>Ipomoea dichroa</i> Hochst. ex Choisy.	Convolvulaceae	LT	SZ	Pal
<i>Khaya senegalensis</i> (Des.) A. Juss.	Meliaceae	mP	SZ	A
<i>Kigelia africana</i> (Lam.) Benth.	Bignoniaceae	mP	GC-SZ	A
<i>Lagenaria siceraria</i> (Molina) Standl.	Cucurbitaceae	Lmp	GC-SZ- Sah.S	Pt
<i>Lannea acida</i> A. Rich.	Anacardiaceae	mp	GC-SZ	A
<i>Lannea microcarpa</i> Engl. et K. Krauze	Anacardiaceae	mp	SZ	A
<i>Launaea chevalieri</i> O. Hoffm. et Musch.	Asteraceae	T	SZ	A
<i>Lepidagathis anobrya</i> Nees.	Acanthaceae	np	SZ	A
<i>Leptadenia hastata</i> (Pers.) Decne.	Asclepiadaceae	Lmp	SZ-Sah.S	A
<i>Leptadenia pyrotechnica</i> (Forssk.) Decne	Asclepiadaceae	mp	SZ-Sah.S	Pal
<i>Limeum pterocarpum</i> (Gay.) Heimerl.	Aizoaceae	T	SZ-Sah.S	A
<i>Loeseneriella africana</i> (Willd.) Wilczek	Hyppocrateaceae	Lmp	GC-SZ	Pal

Species	Famille	Life	Chorology	
<i>Maerua crassifolia</i> Forssk.	Capparaceae	mp	SZ-Sah.S	A
<i>Maeurua angolensis</i> DC.	Capparaceae	mp	SZ	A
<i>Merremia tridentate</i> (L.) Hallier F.	Convolvulaceae	LT	GC-SZ	A
<i>Mitracarpus scaber</i> Zucc	Rubiaceae	T	GC-SZ	A
<i>Mitragyna inermis</i> (Willd.) K.Schum.	Rubiaceae	mp	SZ	A
<i>Momordica balsamina</i> L.	Cucurbitaceae	LT	SZ-Sah.S	Pt
<i>Neocarya macrophylla</i> (Sabine) Prance	Chrysobalanaceae	mp	GC-SZ	A
<i>Neptunia oleracea</i> Lour.	Elatinaceae	Hy	GC-SZ	Pt
<i>Nymphaea lotus</i> L.	Nymphaeaceae	Hy	GC-SZ	A
<i>Ocimum gratissimum</i> L.	Lamiaceae	T	GC-SZ	Pal
<i>Oncoba spinosa</i> Forssk	Salicaceae	mp	GC-SZ	A
<i>Ozoroa insignis</i> Del.	Anacardiaceae	mp	SZ	A
<i>Parkia biglobosa</i> (Jacq.) R.Br. ex G.Don	Mimosaceae	mp	SZ	A
<i>Paullinia pinnata</i> L.	Sapindaceae	Lmp	GC-SZ	AN
<i>Pavetta crassipes</i> K.Schum.	Rubiaceae	mp	SZ	A
<i>Pennisetum americanum</i> (L.) Leeke (use <i>Pennisetum glaucum</i>)	Poaceae	T	SZ-Sah.S	A
<i>Pennisetum pedicellatum</i> Trin.	Poaceae	T	GC-SZ- Sah.S	Pal
<i>Pergularia daemia</i> (Forssk.) Chiov.	Asclepiadaceae	np	SZ-Sah.S	Pal
<i>Phyllanthus pentandrus</i> Schumach. & Thonn.	Euphorbiaceae	T	GC-SZ	A
<i>Piliostigma reticulatum</i> (DC.) Hochst.	Caesalpiniaceae	mp	SZ-Sah.S	A
<i>Pistia stratiotes</i> L.	Araceae	Hy	GC-SZ	Pt
<i>Polycarpha linearifolia</i> (DC.) DC.	Caryophyllaceae	T	SZ-Sah.S	A
<i>Prosopis africana</i> (Guill. & Perr.) Taub.	Mimosaceae	mP	SZ	A
<i>Pseudocedrela kotschy</i> (Schweinf.) Harms	Meliaceae	mp	SZ	A
<i>Pteleopsis suberosa</i> Engl. & Diels	Combretaceae	mp	SZ	A
<i>Pterocarpus erinaceus</i> Poir.	Fabaceae	mP	SZ	A
<i>Pupalia lappacea</i> (L.) A.Juss.	Amaranthaceae	T	GC-SZ- Sah.S	Pal
<i>Ricinus communis</i> L.	Euphorbiaceae	mp	i	Pal
<i>Rogeria adenophylla</i> J.Gay ex Delile	Pedaliaceae	T	SZ-Sah.S	A
<i>Salvadora persica</i> L.	Salvadoraceae	mp	SZ-Sah.S	Pal
<i>Sansevieria liberica</i> Gérôme & Labroy	Agavaceae	Gr	GC-SZ	A
<i>Sarcocephalus latifolius</i> (Sm.) E.A.Bruce	Rubiaceae	mp	GC-SZ	A
<i>Sarcostemma viminalis</i> (L.) R.Br.	Asclepiadaceae	CH		
<i>Schwenkia americana</i> L.	Solanaceae	T	GC-SZ	AN
<i>Sclerocarya birrea</i> (A.Rich.) Hochst	Anacardiaceae	mP	SZ-Sah.S	A
<i>Scoparia dulcis</i> L.	Scrophulariaceae	np	GC-SZ- Sah.S	Pt
<i>Securidaca longipedunculata</i> Fresen.	Polygalaceae	mp	SZ	A
<i>Sesamum alatum</i> Thonn.	Pedaliaceae	T	SZ-Sah.S	A
<i>Sesamum indicum</i> L.	Pedaliaceae	T	GC-SZ	Pt
<i>Sesbania pachycarpa</i> DC.	Fabaceae	T	SZ	AM
<i>Sida ovata</i> Forssk.	Malvaceae	CH	SZ	Pt

Species	Famille	Life	Chorology	
<i>Solanum incanum</i> L.	Solanaceae	np	SZ-Sah.S	Pal
<i>Solanum lycopersicum</i> L.	Solanaceae	T	GC-SZ- Sah.S	N
<i>Solenostemma argel</i> (Delile) Hayne	Asclepiadaceae	np	Sah.S	A
<i>Sterculia setigera</i> Forssk.	Sterculiaceae	mp	SZ	A
<i>Stereospermum kunthianum</i> Cham.	Bignoniaceae	mp	SZ	A
<i>Striga hermonthica</i> (witchweed)	Scrophulariaceae	T	SZ	AM
<i>Strychnos innocua</i> Delile	Loganiaceae	mp	SZ	A
<i>Stylosanthes erecta</i> P.Beauv.	Fabaceae	T	GC-SZ	A
<i>Taccazea apiculata</i> Oliv.	Periplocaceae	Lmp	GC-SZ	A
<i>Tamarindus indica</i> L.	Caesalpiniaceae	mP	GC-SZ- Sah.S	Pt
<i>Tapinanthus globiferus</i> (A.Rich.) Tiegh.	Loranthaceae	mp	SZ-Sah.S	A
<i>Tephrosia linearis</i> (Willd.) Pers.	Fabaceae	T	SZ	A
<i>Tephrosia lupunifolia</i> DC.	Fabaceae	CH	SZ	A
<i>Tephrosia obcordata</i> (Lam. ex Poir.) Baker	Fabaceae	CH	SZ	As
<i>Tephrosia purpurea</i> (L.) Pers.	Fabaceae	CH	SZ-Sah.S	Pal
<i>Terminalia avicennioides</i> Guill. & Perr.	Combretaceae	mP	SZ	A
<i>Tribulus terrestris</i> L.	Zygophyllaceae	T	GC-SZ	Pt
<i>Vernonia colorata</i> (Willd.) Drake	Asteraceae	np	GC-SZ	A
<i>Vigna unguiculata</i> (L.) Walp	Fabaceae	LT	GC-SZ	Pt
<i>Vitellaria paradoxa</i> C.F.Gaertn.	Sapotaceae	mp	SZ	A
<i>Vitex doniana</i> Sweet	Lamiaceae	mp	GC-SZ	A
<i>Waltheria indica</i> L.	Sterculiaceae	np	GC-SZ	Pt
<i>Ximenia americana</i> L.	Olacaceae	mp	GC-SZ	Pt
<i>Ziziphus mauritiana</i> Lam.	Rhamnaceae	mp	SZ-Sah.S	A
<i>Ziziphus mucronata</i> Willd.	Rhamnaceae	Lmp	SZ	AM
<i>Zornia glauchidiata</i> Rchb. ex DC	Fabaceae	T	GC-SZ	A

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