



# Microorganisms and Carrier Molecules used in Biofertilizer Formulations

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## Authors' contributions

*This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.*

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## ABSTRACT

Rhizobacteria with plant growth promoting potential are used widely for agricultural use. They have been used as supplements to chemical fertilizers. Cyanobacteria, nitrogen fixing bacteria, phosphate mineralizing bacteria, and potassium solubilizing microorganisms have been used as biofertilizers for different crops. The survival of these rhizobacteria in soil is affected by number of abiotic as well as biotic factors that limits the growth of rhizobacteria. To enhance the survival rate of rhizobacteria in soil and to enhance their viability and efficiency, the bacteria is mixed with carrier material or additive to increase their activity. Carrier molecule enhances the adhesion of bacteria to the plant roots. The processing of bacteria with carrier molecule ensures easy handling and processing or agricultural use. Addition of additive molecule to the bacterial culture provides excellent survival rate and slow cell release at different soil pH. Different additive molecules used as carrier molecules are discussed in this article.

*Keywords: Biofertilizers; inoculants; carrier molecule; additive; growth; development.*

## 1. INTRODUCTION

Biofertilizers are "Preparations containing live microbes that augment the plant growth by enhancing nutrient availability in soil and

protecting the crop from various biotic and abiotic factors [1]. Biofertilizers holds great potential in sustainable agriculture. They can be used as a replacement to chemical fertilizers in organic farming and forms an important part of integrated

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nutrient management system. Microorganisms including phosphate and potassium solubilizing fungi/bacteria, nitrogen-fixers, cyanobacteria and other microorganisms with plant growth promoting traits can be utilized as biofertilizers [2]. One of the most attractive features of biofertilizers is their environment friendly and cost-effective nature.

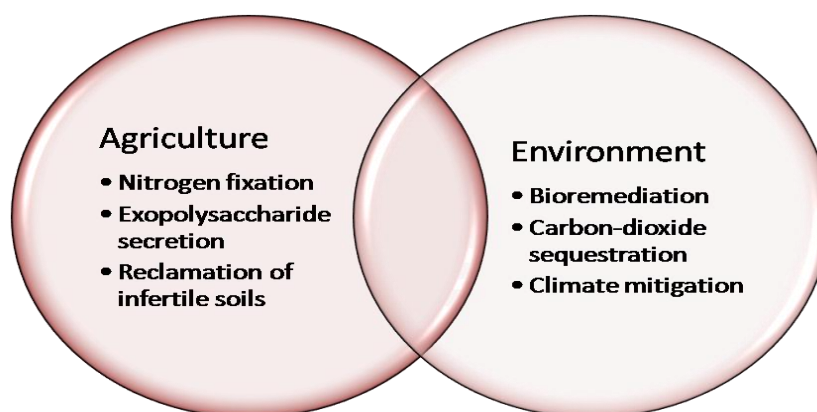
Different micro inoculums are used as biofertilizer for good practices of agriculture for increasing crop yield. These microorganisms themselves contain various properties which are vital for increasing the product value. The microbial formulations are used to stimulate certain processes which can convert insoluble nutrients to soluble forms that can be used by plants. The microbial inoculum is mixed with carrier molecule that helps the microbial culture to bind to it and maintain its viability for longer period. A carrier is basically a formulation carrying microorganisms, peat, vermiculite, lignin, etc. [3]. Different microorganisms that can be used as biofertilizer are discussed below-

### 1.1 Cyanobacteria

Cyanobacteria (blue green algae) are abundantly found in marine environment and have a great potential to be used for sustainable agriculture [4]. It is well known to maintain the nitrogen level in soil, improve its water holding capacity and aeration [5]. *Nostoc* and *Anaebena* are two most common cyanobacteria used as biofertilizer (Table 1). The striking feature of cyanobacteria that makes it suitable for use as biofertilizer is that it does not require any kind of substructure or host for its cultivation, growth, and production of molecules responsible for plant growth promoting. *Azolla-Anaebena* that is known for its symbiotic association also serves to increase crop yield. Cyanobacterial biofertilizers can be directly supplied into the soil or can be applied to the seeds. Due to the exopolysaccharide secreting potential, the cyanobacteria can be used for the reclamation of infertile soils [6] They also have roles in reducing global warming and maintaining bio-geochemical cycles (Fig. 1).

**Table 1. Effect of cyanobacteria on different crop**

Bacterial species	Crop Effect	Effect on the crops	Reference
<i>NostocPunctiforme</i> PC C 73102	Rice ( <i>Oryza sativa</i> L.)	Yield of plant was increased	Álvarez et al. [7]
Cyanobacteria	Lettuce ( <i>Lactuca sativa</i> L)	Leaf length, fresh and dry weight of the plant was increased	Menamo &Wolde [8]
Cyanobacterium <i>Anaba ena</i>	Wheat	Phosphorous uptake was enhanced	Swarnalakshmi et al., [9]
<i>Nostoc</i>	Maize	Enhances structure of soil, soil fertility, and growth of maize	Maqubela et al., [10]



**Fig. 1. Different roles of cyanobacteria [4]**

## 1.2 Nitrogen Fixing Bacteria

Nitrogen is the macro nutrient essential for the growth and development of non-legume plants and cereals. Therefore, nitrogen must be supplied through fertilizers. However, injudicious use of synthetic nitrogenous fertilizers contributes to contamination of soil and groundwater, leading to human health hazards, and threatens agricultural sustainability [3]. Rhizobium containing biofertilizer enhance the productivity of leguminous plants through supply of nitrogen via symbiotic associations. These bacteria induce nodule formation in leguminous plants. Various other bacteria belonging to the genus *Azotobacter*, *Azoarcus*, *Burkholderia*, *Enterobacter*, *Gluconacetobacter*, *Azospirillum*, *Pseudomonas* have potential to fix nitrogen non-symbiotically in wheat, rice, sunflower, maize and other non-legume crops [11] (Tables 2,3).

## 1.3 Actinomycetes

Actinomycetes comprise a major group of microorganisms which are found in rhizosphere and inside the plant roots as endophytes and are documented for their role to improve plant growth. They are well known for their potential to

recycle nutrient by degradation of chitin, cellulose, starch, lipids and complex carbohydrates and converting them into simple sugars. Actinomycetes are the key component of agricultural ecosystems and are important in promotion of plant growth by various means (Fig. 2). These microorganisms around the plant root surfaces perform an important role of breakdown of organic matter and making it available for the plant uptake. These microorganisms also show their potential role in solubilization of phosphate, production of siderophores, hydrogen cyanide, auxin, ammonia and lytic enzymes [16]. Their morphology renders them more efficient for solubilizing phosphorus by increasing surface to volume ratio. Actinomycetes are known to secrete various enzymes and low molecular weight organic acids that help in solubilization of phosphate and potash rocks. Some species of actinomycetes such as *Streptomyces venezuelae*, *S. alboniger*, *S. ambofaciens* and *S. lienomycini* have been known to secrete phytate degrading enzymes phytase that help in solubilization of organic phosphorus pool in the soil and making it available to the plants. The potential of actinomycetes to produce indole acetic acid also makes them important candidate for use as biofertilizers [17].

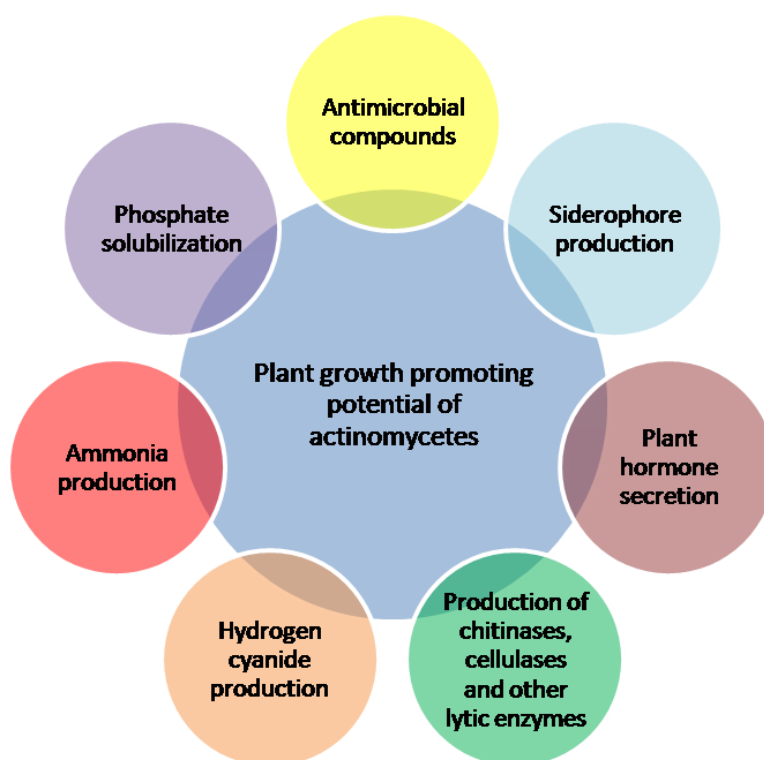


Fig. 2. Different bioactive produced by actinomycetes [16]

**Table 2. Effect of nitrogen fixing bacteria on different crop**

<b>Bacterial species</b>	<b>Crop</b>	<b>Effect on the crops</b>	<b>Reference</b>
<i>Azospirillum lipoferum</i> or <i>Azotobacter chroococcum</i>	Maize	It helps in maintaining the negative effects on salt stress on maize plants. Improves physiological activities under saline conditions for maize growth.	Latef et al., [12]
<i>Azospirillum brasilense</i>	Maize	Enhanced growth of maize. Nitrogen use efficiency of maize was also increased.	Zeffa et al., [13]
<i>Rhizobium leguminosarum</i>	Lettuce ( <i>Lactuca sativa</i> ) and Carrots ( <i>Daucus carota</i> )	Dry matter weight was increased. Increases uptake of N and P in the edible parts of both plants.	Flores-Félix et al. [14]
<i>Rhizobium Leguminosarum</i> (RhOF34, RhOF125 and RhOF15)	Broad bean, Fava bean, (VICIA FABA)	Improved growth of the plants. Mineral and nitrogen uptake was improved.	Benidire, [11]
<i>Pseudomonas stutzeri</i> A1501	Rice	It helps in the growth of rice in the presence of salt concentration and heavy metals.	Han, [15]

**Table 3. Effect of nitrogen fixing bacteria on different crop**

<b>Bacteria species</b>	<b>Crop</b>	<b>Effects on the crop</b>	<b>Reference</b>
Bacillus spp.	Crop plants (sugar cane, wheat, rice, corn, maize)	Helped managing the abiotic stresses (water, salinity and heavy metal).	Nanjundappa [18]
<i>Micrococcus sp.</i> TISTR2221	Maize grain ( <i>Z. mays</i> L.)	Significantly promoted shoot length, root length, and dry biomass of Maize.	Sangthong et al., [19]
<i>Flavobacterium johnsoniae</i> strain GSE09	Black Pepper ( <i>Piper nigrum</i> )	Exhibited biofilm formation and produced indolic compounds and showed biocontrol activity against <i>Phytophthora capsici</i> .	Sang et al., [20]
<i>Aspergillus niger</i> F7	Crop plants (sugar cane, wheat, rice, corn, maize)	Maintained available phosphate level for crops.	Srividya et al., [21]
<i>Penicillium funiculosum</i> LHL06	Soybean ( <i>Glycine max</i> L.)	Improved soybean crop growth by reducing toxic effects on metals. Gibberellin production was observed during copper stress.	Khan & Lee [22]
<i>Pseudomonas fluorescens</i>	Crop plants (sugar cane, wheat, rice, corn, maize)	Known to produce antibiotics, volatiles, siderophores, and growth-promoting substances	Khan & Lee [22]

Bacteria species	Crop	Effects on the crop	Reference
<i>Pseudomonas fluorescens</i> (SS5)	Tomato	Plant growth promotion reported due to increase in root, shoot weight, length and fruit yield per plant.	Ahirwar et al., [23]

## 2. CARRIER AND ADDITIVES FOR BIOFERTILIZER

Plant growth promoting microorganisms are served as bioinoculants for agricultural use by incorporating them in carrier material for easy handling, high effectiveness and high shelf life. Various different types of carrier materials are used for biofertilizer development. The carrier material should have certain properties such as:

1. Non-toxic to inoculant bacterial strain
2. Good moisture absorption capacity
3. Easy to process and free of lump-forming materials
4. Easy to sterilize
5. Available in adequate amounts
6. Inexpensive
7. Good adhesion properties
8. non-toxic to plant

For soil inoculation, carrier material like peat, charcoal, perlite, clay of size 0.5-1.5mm is generally used. For seed inoculation, the carrier material should be milled to fine powder with particle size of 10 -40 µm.

### 2.1 Clay Minerals as Additives

There are some wide ranges of carrier material used in bio immobilized system which provides various properties such as bulking capacity, stability and protection against physical stress. To release microorganism slowly into the soil and to protect the microbial cells against desiccation, the combinations of soil microorganism inoculants and clay minerals is used [24]. This combination had provided a great effect on the significantly increasing microorganism survival by reducing UV effects compared to normal cells [25]. For example, when *Pseudomonas* microencapsulated in a mixture of alginate and starch was combined, under saline conditions there was increase in soluble protein content, carotenoid concentrations and cotton biomass, chlorophylls [26]. Clay minerals such as pyrophyllite, bentonite and kaolin in combination with alginate, is used for immobilization of *Pantoea agglomerans*, *Trichoderma*

*harzianum* and other plant growth promoting bacteria [27].

### 2.2 Skim Milk

Skimmed milk is used in bio formulations which enhances cell viability during storage conditions [28]. Encapsulation of *Azospirillum* and *Pseudomonas* with skim milk powder significantly increases the cell number and viability [24]. The cell release rate is also found to higher in cells encapsulated with skim milk. The recovery rate of 100% in *P. fluorescens* was observed in alginate- skim milk encapsulated beads. For encapsulation of rhizobacteria, skim milk can be used in combination of clay mineral for strategic increase in cell count and cell viability.

### 2.3 Starch

Starch is used as carrier or additive for use in encapsulation of rhizobacteria. Starch is reported to improve the survival of bacterial cells by reducing the stress on microbial cells [24]. The microbial cells adhere to the starch molecules which protects it from physical stress.

The cell adhesion to starch is based on the effect of starch and stress conditions on the microbial cells.

### 2.4 Chitin/Chitosan as Additives

Chitin or its polymer chitosan, is a bioactive oligosaccharide polymer which is used as filler material or a cell protectant in biofertilizers. Chitin is known for its antibacterial activity, non-toxic and biodegradability potential [29]. Reports have shown that the encapsulation of *Bacillus* in a matrix containing chitin, improves the multiplication of bacterial cells and enhances its antifungal potential [30]. Also, for the biocontrol activity, chitin is an excellent chelating agent against pathogens [31] and it also enhances stress tolerance, antioxidant activity, and osmoregulator potential in plants [32]. Being a coating material, it plays an important role in lowering the formulation cost and providing the strength to plants. Chitosan is an excellent

carrier which can maintain the viability of beneficial microorganisms for plants [33]. Chitosan has also been widely used for seed coating. Seed of soyabean, maize, wheat, rice and peanut were found to have better germination rates when coated with chitin [34].

## 2.5 Humic Acid Additives

To promote the populations of specific microorganism, humic acid and its derivatives are used in biofertilizers [35]. Formulation of microbial inoculants immobilized with humic acid additives has shown excellent survival, releases cell slowly at various pH, and provide uniform growth to bacteria. To enhance root colonization by fungi, humic acid is usually added in biofertilizers [36,37]

## 3. CONCLUSION

The development of microbial bioformulations for agricultural use has leads us towards the sustainable option against chemical pesticides and fertilizers. Towards this approach, microbial inoculants with plant growth promoting activities are being used. However, their direct application in soil decreases their viability and plant growth promoting potential. Therefore, they are applied in combination with carrier molecules or additives that help the bacteria binds to it and enhances its survival rate in the soil. Different carrier molecules are used which have been discussed. Currently, the carrier based biofertilizers are being replaced by encapsulated biofertilizers. These additives can also be used as polymer matrix for encapsulation.

## CONSENT

It is not applicable.

## ETHICAL APPROVAL

It is not applicable.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Kaur R, Kaur S. Variation in the phosphate solubilizing bacteria from virgin and the agricultural soils of Punjab. *Cur Microbiol*, 2020;77(9):2118-2127.

2. Rani N, Kaur R, Kaur S. Zinc solubilizing bacteria to augment soil fertility: A comprehensive review. *Int Agri Sci Vet Med*. 2020;8(1):38-44.
3. Kaur R, Kaur S. Biological alternates to synthetic fertilizers: efficiency and future scopes. *Indian Journal of Agricultural Research*. 2018;52(6).
4. Pisciotta JM, Zaybak Z, Call DF, Nam JY, Logan BE. Enrichment of microbial electrolysis cell biocathodes from sediment microbial fuel cell bioanodes. *Applied and Environmental Microbiology*. 2012;78(15):5212-5219.
5. Paumann M, Regelsberger G, Obinger C, Peschek GA. The bioenergetics role of dioxygen and the terminal oxidase(s) in cyanobacteria, *Biochim. Biophys. Acta Bioenerg*. 2005;1707.
6. Paul D, Nair S. Stress adaptations in a plant growth promoting rhizobacterium (PGPR) with increasing salinity in the coastal agricultural soils. *J. Basic Microbiol*. 2008;48:378–384. DOI: 10.1002/jobm.200700365
7. Álvarez C, Navarro JA, Molina-Heredia FP, Mariscal V. Endophytic colonization of rice (*Oryza sativa* L.) by the symbiotic strain *Nostoc punctiforme* PCC 73102. *Molecular Plant-Microbe Interactions*. 2020;33(8):1040-1045.
8. Menamo M, Wolde Z. Effect of cyanobacteria application as biofertilizer on growth, yield and yield components of romaine lettuce (*Lactuca sativa* L.) on Soils of Ethiopia. *American Scientific Research Journal for Engineering, Technology, and Sciences (ASRJETS)*. 2013; 4(1):50-58.
9. Swarnalakshmi K, Prasanna R, Kumar A, Pattnaik S, Chakravarty K, Shivay YS, Saxena AK. Evaluating the influence of novel cyanobacterial biofilmed biofertilizers on soil fertility and plant nutrition in wheat. *European Journal of Soil Biology*. 2013;55:107-116.
10. Maqubela MP, Mnkeni PNS, Issa OM, Pardo MT, D'acqui LP. *Nostoc* cyanobacterial inoculation in South African agricultural soils enhances soil structure, fertility, and maize growth. *Plant and Soil*. 2009;315(1):79-92.
11. Benidire L, Lahrouni M, El Khalloufi F, Gottfert M, Oufdou K. Effects of *Rhizobium leguminosarum* inoculation on growth, nitrogen uptake and mineral assimilation in *Vicia faba* plants under salinity stress; 2017.

12. Latef AAHA, Alhmad MFA, Kordrostami M, Abo-Baker ABAE, Zakir A. Inoculation with *Azospirillum lipoferum* or *Azotobacter chroococcum* reinforces maize growth by improving physiological activities under saline conditions. *Journal of Plant Growth Regulation*. 2020;39(3):1293-1306.
13. Zeffa D.M, Perini LJ, Silva MB, de Sousa NV, Scapim CA, Oliveira ALMD, Azeredo Goncalves LS. *Azospirillum brasilense* promotes increases in growth and nitrogen use efficiency of maize genotypes. *PLoS One*. 2019;14(4):e0215332.
14. Flores- Félix JD, Menéndez E, Rivera LP, Marcos- García M, Martínez- Hidalgo P, Mateos PF, Rivas R. Use of *Rhizobium leguminosarum* as a potential biofertilizer for *Lactuca sativa* and *Daucus carota* crops. *Journal of Plant Nutrition and Soil Science*. 2013;176(6):876-882.
15. Han Y, Wang R, Yang Z, Zhan Y, Ma Y, Ping S, Yan Y. 1-aminocyclopropane-1-carboxylate deaminase from *Pseudomonas stutzeri* A1501 facilitates the growth of rice in the presence of salt or heavy metals. *Journal of Microbiology and Biotechnology*. 2015;25(7):1119-1128.
16. Sousa JAJ, Olivares FL. Plant growth promotion by streptomycetes: ecophysiology, mechanisms and applications. *Chem*; 2016
17. Rani K, Dahiya A, Masih JC, Wati L. Actinobacterial biofertilizers: an alternative strategy for plant growth promotion. *Int J Curr Microbiol App Sci*. 2018;7(09):607-614.
18. Nanjundappa A, Bagyaraj DJ, Saxena AK, Kumar M, Chakdar H. Interaction between arbuscular mycorrhizal fungi and *Bacillus* spp. in soil enhancing growth of crop plants. *Fungal Biology and Biotechnology*. 2019;6(1):1-10.
19. Sangthong C, Setkit K, Prapagdee B. Improvement of cadmium phytoremediation after soil inoculation with a cadmium-resistant *Micrococcus* sp. *Environmental Science and Pollution Research*. 2016;23(1):756-764.
20. Sang MK, Kim KD. The volatile- producing *Flavobacterium johnsoniae* strain GSE09 shows biocontrol activity against *Phytophthora capsici* in pepper. *Journal of Applied Microbiology*. 2012;113(2):383-398.
21. Srividya S, Soumya S, Pooja K. Influence of environmental factors and salinity on phosphate solubilization by a newly isolated *Aspergillus niger* F7 from agricultural soil. *African Journal of Biotechnology*. 2009;8(9).
22. Khan AL, Lee IJ. Endophytic *Penicillium funiculosum* LHL06 secretes gibberellin that reprograms *Glycine max* L. growth during copper stress. *BMC Plant Biology*. 2013;13(1):1-14.
23. Ahirwar NK, Gupta G, Singh V, Rawley, RK, Ramana S. Influence on growth and fruit yield of tomato (*Lycopersicon esculentum* Mill.) plants by inoculation with *Pseudomonas fluorescence* (SS5): Possible role of plant growth promotion. *Int. J. Curr. Microbiol. Appl. Sci*. 2015; 4(2):720-730.
24. Bashan Y, Hernandez JP, Leyva LA, Bacilio M. Alginate microbeads as inoculant carriers for plant growth-promoting bacteria. *Biol. Fertil. Soils*. 2002;35:359-368.
25. He Y, Wu Z, Tu L, Han Y, Znahg G, Li C. Encapsulation and characterization of slow-release microbial fertilizer from the composites of bentonite and alginate. *Appl. Clay Sci*. 2015;109-110:68-75.
26. He Y, Wu Z, Tu L, Shan C. Effect of encapsulated *Pseudomonas putida* Rs-198 strain on alleviating salt stress of cotton. *J. Plant Nutr*. 2017;40:1180-1189.
27. Zohar-Perez C, Chernin, L, Chet, I, and Nussinovitch, A. Structure of dried cellular alginate matrix containing fillers provides extra protection for microorganisms against UVC radiation. *Radiat. Res*. 2003; 160:198-204. DOI: 10.1667/rr3027.
28. Yu WK, Yim TB, Lee KY, Heo TR. Effect of skim milk-alginate beads on survival rate of *Bifidobacteria*. *Biotechnol. Bioprocess Eng*. 2001;6:133-138.
29. Muxika A, Etxabide A, Uranga J, Guerrero P, de la Caba K. Chitosan as a bioactive polymer: processing, properties and applications. *Int.J. Biol. Macromol*. 2017; 105:1358-1368.
30. Manjula K, Podile AR. Chitin supplemented formulations improve biocontrol and plant growth-promoting efficiency of *Bacillus subtilis* AF1. *Can. J. Microbiol*. 2001; 47:618-625.
31. Berger LRR, Stamford TCM, Stamford-Arnoud, T. M, Alcântara SRC, Silva, A. C, Silva, A. M, et al. Green conversion of agroindustrial wastes into chitin and chitosan by *Rhizopus arrhizus* and *Cunninghamella elegans* strain. *Molecules* 2014;15:9082-9102.

32. Dar TA, Uddin M, Khan MM, Ali A, Mir, SA, Varshney, L. Effect of Co-60 gamma irradiated chitosan and phosphorus fertilizer on growth, yield and trigonelline content of *Trigonella foenum-graecum* L. J. Radiat. Res. Appl. Sci. 2015;8:446–458.
33. Chanratana, M, Han, G. H, Joe, M. M, Choudhury, A. R, Sundaram, S, and AbdulHalim, MDet al. Evaluation of chitosan and alginate immobilized *Methylobacteriumoryzae* CBMB20 on tomato plant growth. Arch. Agron. Soil Sci. 2018;64:1489–1502.
34. Crini, M, N, Lichtfouse, E, Torri, G, &Crini, G. Applications of chitosan in food, pharmaceuticals, medicine, cosmetics, agriculture, textiles, pulp and paper, biotechnology, and environmental chemistry. Environmental Chemistry Letters. 2019;17(4):1667-1692.
35. Pukalchik, M, Kydralieva, K, Yakimenko, O, Fedoseeva, E, and Terekhova, V. Outlining the potential role of humic products in modifying biological properties of the soil—a review. Front. Environ. Sci. 2019;7:80.
36. Gryndler, M, Larsen, J, Hrselova, H, Rezacova, V, Gryndlerova, H, and Kubat, J. Organic and mineral fertilization, respectively, increase and decrease the development of external mycelium of arbuscular mycorrhizal fungi in a longterm field experiment. Mycorrhiza. 2005;16: 159–166.
37. Meeks JC, Campbell EL, Summers ML, Wong FC. Cellular differentiation in the cyanobacterium *Nostocpunctiforme*. Archives of Microbiology. 2002;178(6):395-403.

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